HOSTED BY

ELSEVIER

Contents lists available at ScienceDirect

Electronic Journal of Biotechnology



Polymorphism of three milk protein genes in Mexican Jersey cattle



José Luis Zepeda-Batista, Baldomero Alarcón-Zúñiga, Agustín Ruíz-Flores *, Rafael Núñez-Domínguez, Rodolfo Ramírez-Valverde

Posgrado en Producción Animal, Departamento de Zootecnia, Universidad Autónoma Chapingo, Km 38.5 Carretera México-Texcoco, Edo. de México CP 56230, México

ARTICLE INFO

Article history:
Received 12 August 2014
Accepted 3 October 2014
Available online 10 November 2014

Keywords: β-Casein β-Lactoglobulin Genetic diversity κ-Casein

ABSTRACT

The objective was to estimate the allelic and genotypic frequencies, genetic diversity and polymorphic information content for the β -casein, κ -casein and β -lactoglobulin genes. Blood and frozen semen samples were collected from 453 Jersey individuals registered by the Mexican Jersey Cattle Association. Twenty eight breed specific SNP primers for whole genes were used. The B allele of κ -casein had higher frequency (0.69) than the A (0.26) and E (0.05). For β -lactoglobulin, the highest frequency was for B (0.72), followed by A and C alleles (0.26 and 0.02, respectively). The β -casein allele with the highest frequency was A^2 (0.71), followed by A^1 (0.19), A^3 (0.05), B (0.04) and C (0.01). The average genetic diversity (He) was 0.53. The average locus effective allele number was 1.79. These results indicate a high allelic diversity for κ -caseín, β -casein and β -lactoglobulin that could be included in breeding programs in the population studied, aimed to improve the milk quality traits of economic importance.

© 2014 Pontificia Universidad Católica de Valparaíso. Production and hosting by Elsevier B.V. All rights reserved.

1. Introduction

Mexican dairy cattle populations have been developed from imported genetic resources worldwide, incorporating a wide base of milk protein genes, including $\alpha S1$ -casein, $\alpha S2$ -casein, β -casein, κ -casein, α -lactoalbumin and β -lactoglobulin. Particularly, the Jersey cattle populations have been constituted by American, Canadian [1], European, Australian and New Zealander progenitors. Jersey populations of these countries have been reported with variable gene frequencies for milk protein genes [2,3,4].

The importance to detect the genetic polymorphism for milk protein genes in dairy cattle populations is their association with cheese yield, rennet time, and curd firmness [3,5,6]. Besides, most of the available technologies have breed specific developments; therefore, when this technology is applied on breeds different than the one used as a model, imprecisions at the presence of genetic marker level from 8.3 to 54.9% might appear [7,8]. Molecular technologies have been developed to detect alleles and frequencies within protein milk genes, including specific PCR sequences, restriction enzymes, and actually single nucleotide polymorphism [9,10,11]. The development of breed specific SNPs is necessary for genotyping and association mapping to milk traits. The objectives were to determine the allelic and genotypic frequencies, genetic diversity and polymorphic information

E-mail addresses: arf@correo.chapingo.mx, aruizf1959@hotmail.com (A. Ruíz-Flores). Peer review under responsibility of Pontificia Universidad Católica de Valparaíso.

for $\beta\text{-casein},$ $\kappa\text{-casein},$ and $\beta\text{-lactoglobulin}$ in Mexican Jersey cattle populations.

2. Materials and methods

2.1. Samples for DNA extraction

Samples were collected from 453 Jersey individuals registered by the Mexican Jersey Cattle Association, originated from Canadian, U.S., New Zealand, Australian and Mexican progenitors, including 401 cows and 52 sires. Sampled cows had at least a calf, whereas sires needed to have at least two calves in different herds. DNA samples from cows were obtained from blood and frozen semen was used to obtain the DNA in sires.

2.2. Selection of SNP primers

Primers used to genotype individuals were designed using OligoAnalyzer 3.1® (Integrated DNA Technologies, 2012, Iowa, USA), corresponding to the exons that represent the open reading frame of β-casein (β-CSN-encoding gene CSN2), κ-casein (κ-CSN-encoding gene CSN3), and β-lactoglobulin (β-LGB-encoding gene LGB) milk proteins reported in GenBank (NC_007304.4, http://www.ncbi.nlm.nih.gov/genbank; [12]). The changing nucleotides were marked with a different fluorophore at the SNP position to distinguish each one during the allele identification by real time PCR. The reverse sequences and their complements, the coefficients of hairpin formation, autodimerization and creation of heterodimers, the

^{*} Corresponding author.

percentage of each nucleotide, and the fusion point for each sequence, necessary to design the thermo-cycling protocol, were determined with the same software.

Total SNP primers for whole genes were 28, including 8 primer sequences for β -lactoglobulin, 10 for κ -casein and 10 for β -casein. Primers were synthesized by KBioscience (Massachusetts, USA).

Molecular validation was done by amplifying the previously designed primers to corroborate the in silico performance. With the SNP primers that amplified well, the resolution curve was verified to guarantee the correct measure of the sample.

2.3. Genotyping of DNA

The PCR amplification was done with 4.11 µL of reaction solution (KASPar V4.0 Master Mix, KBioscience, USA) and 20 ng of genomic DNA; for the negative template control, 4 µL of molecular grade water was used. The polymorphisms were identified with the UV–Vis of the rtPCR thermo cycler (CFX 96, Bio–Rad, California, USA). Different fluorescence of SNP primers were used to detect the changing nucleotides. The thermocycling protocol was: one cycle at 94°C for 15 min, 10 cycles at 94°C for 20 s and 65°C for a min, with a decrease of 0.8°C per cycle in the second step, and 35 cycles at 94°C for 20 s, followed by a cycle at 57°C for 20 s, including the fluorescence report for each cycle in the last step. With the 28 validated SNPs, it was possible to identify the most common alleles for the three milk proteins. This increased the probability of detection of those alleles previously reported as being of low frequency in *Bos taurus* populations [13].

2.4. Data analysis

Polymorphic amplicons were considered to estimate the allelic diversity and effective number of alleles. Allelic and genotypic frequencies were estimated using the software Popgen32 [14].

3. Results and discussion

3.1. Genotyping for the three milk proteins

Data of haplotype and genotype frequencies of three protein milk genes in 453 individuals representing the Mexican Jersey cattle gene pool, are reported.

3.1.1. β-Casein

The alleles identified were A^1 , A^2 , A^3 , B and C (Table 1); A^2 was the most frequent in the Mexican Jersey populations, 0.71. The frequency for this allele was similar to those reported, 0.58 to 0.65, for other Jersey populations [15,16,17]. This result suggests an absence of genetic selection for β -casein in Jersey cattle, including the Mexican populations. Some researchers have shown that the presence of the A^2 allele in dairy cattle produces high quality milk associated with diminished cholesterol and triglycerides in humans [18,19]. The fact that A^2 allele and A^2A^2 genotype were high in the Mexican Jersey populations is an important and distinctive aspect of this breed that could be used to improve the margin of profit for the milk producers. The frequency for the A^1 allele in this study, 0.19, was similar to that reported by Van Eenennaam and Medrano [2] in a US Brown Swiss

population, 0.18, which also is close to the frequencies for this allele found in Guernsey and Jersey, breeds with high total solids in milk [16,20]. This is in contrast to the frequencies of the A¹ allele reported for Holstein populations, 0.49 to 0.95 [21,22].

The frequency of the A³ allele was 0.05, similar to that reported by Ng-Kwai-Hang and Grosclaude [23], who found a frequency for this allele in US Holstein-Friesian and Brown Swiss of 0.04. Meanwhile, the B allele had low frequencies in the Jersey populations, 0.04, similar to the frequencies reported for Jersey in some American and German populations, as well as for American Brown Swiss [2,20,24]. The low allele frequencies of A³ and B in this study, and the results reported by different authors in dairy breeds, suggest that these polymorphisms have little importance for breed differences in dairy cattle.

The most common genotype for β -casein was A^2A^2 , 0.53. These results differ from those reported by Çardak [25] in a Turkish Holstein-Friesian population where the most frequent genotype was A¹A², 0.46. Furthermore, in this study the genotypes A¹A³, A¹B, and A²C were found, with frequencies lower than 0.05, which are similar to those reported by different authors, from 0.01 to 0.06, in US, New Zealand and Denmark Holstein, Brown Swiss and Ayrshire populations [16,20]. The genotypic frequencies found in the whole population were similar to those estimated only in females (Table 1). However, those corresponding to the male sub-population were different, with some absent genotypes (A^2A^3 , A^1B , A^2C , A^3A^3 , and BB). Hanusová et al. [26] found in Polish Holstein that one of the genotypes did not occur in the male subpopulation, even if it was present in the whole population. The absence of some alleles and genotypes in the male sub-population of this study might be a consequence of the reduced number of sampled individuals (n = 51 sires), even if the origin of sires is diverse.

3.1.2. к-Casein

The alleles detected were A, B, and E (Table 2); B was the most common, 0.69. These findings were similar to the results reported for Colombian, German, and Chinese Jersey populations, 0.71 to 0.89 [11,27,28]. Some authors have shown that the presence of B allele in dairy cattle improves yield and quality of milk, raises milk casein fraction and diminishes whey protein fraction [3,6]. These results suggest an indirect genetic selection for the B allele in Jersey cattle, including the population studied, because the κ-casein alleles related to low total solid production, A (0.26) and E (0.05), are in lower frequencies than B allele. The frequency for the A allele in this population studied was similar to those reported by Trujillo et al. [27] and Ren et al. [11] in Jersey populations in Colombia and China, 0.26 in both populations, as well as in studies with Normande and Guernsey populations [27]. On the contrary, in Holstein there have been reported high frequencies, 0.68 to 0.89, for the A allele in several populations [22,29].

Jann et al. [28] and Boetcher et al. [30] reported in Holstein population frequencies for the E allele of 0.08 and 0.32, respectively. The frequencies for the A and E alleles in the Mexican Jersey populations, could be explained by the objective of selection over time for this breed, total solid production, common among most of the Mexican Jersey based milk producers. This is because these alleles influence positively milk production and negatively total solids in milk, and were indirectly selected against the genetic pool in Jersey.

Table 1 Allelic and genotypic frequencies of β -casein genes in the Mexican Jersey cattle populations.

	n	Allele frequencies					Genotype frequencies									
		A1	A2	А3	В	С	A1A1	A1A2	A1A3	A1B	A2A2	A2A3	A2B	A2C	A3A3	BB
Cows	401	0.22	0.69	0.04	0.04	0.01	0.06	0.30	0.01	0.01	0.50	0.03	0.04	0.01	0.02	0.01
Sires	52	0.12	0.86	0.01	0.01	N/P	0.02	0.20	N/P	N/P	0.73	0.03	0.02	N/P	N/P	N/P
Total	453	0.19	0.71	0.05	0.04	0.01	0.04	0.29	0.01	0.01	0.53	0.03	0.04	0.01	0.02	0.01

Table 2 Allelic and genotypic frequencies in κ-casein gene in the Mexican Jersey cattle populations.

	n	Allele frequencies			Genotype frequencies							
		A	В	Е	AA	AB	AE	BB	BE	EE		
Cows	401	0.27	0.67	0.06	0.04	0.45	0.02	0.42	0.04	0.02		
Sires	52	0.13	0.83	0.04	0.04	0.18	N/P	0.70	0.08	N/P		
Total	453	0.26	0.69	0.05	0.04	0.42	0.02	0.45	0.05	0.02		

The most common genotype frequency for the κ -casein in this study was BB, 0.45. This result differs from those reported by Hernández et al. [31] and Cervantes et al. [32] in Holstein and Criollo Lechero Tropical Mexican populations, where the most frequent genotype was AA, 0.57 and 0.62. Additionally, in the present study the genotypes BE and EE were found, with frequencies lower than 0.1, which were similar to the 0.08 reported by Boetcher et al. [30] for an Italian Holstein population.

3.1.3. \(\beta\)-Lactoglobulin

The alleles found in the sampled cows and sires were A, B, and C (Table 3). Allele B had the highest frequency, 0.72, higher than the estimates reported by Miciński et al. [17], Meza-Nieto et al. [33], and Ren et al. [11] in Holstein, Braunvieh, Jersey, and Criollo Lechero Tropical populations, 0.31 to 0.65. Since the B allele of β -lactoglobulin is related to a high production of total solids, and given its relatively scarce and recent information on the polymorphism for this locus, the differences among the frequencies for this allele in several populations worldwide, suggest selection against it, by the means of selection in favor of the B allele of κ -casein gene in dairy cattle. The frequency observed for the A allele, 0.26, was similar to that reported by Ren et al. [11] in a Chinese Jersey population.

The frequency estimated for the C allele, 0.02, was similar to 0.01 reported by Berry et al. [4] in a New Zealand herd of F1 Holstein x Jersey. The result from the present study could be explained because some of the sampled cows in the Mexican Jersey population were imported from New Zealand.

The most frequent genotype for β -lactoglobulin in the present study was BB, 0.54. This result is similar to the findings of Meza-Nieto et al. [33] in Criollo Lechero Tropical, where the frequency for the BB genotype was 0.53. In Turkish and Mexican Holstein populations, several authors have found frequencies between 0.24 and 0.38 [22,29,33].

Both genotypes BC and AA had a relatively low frequencies, 0.02 and 0.09, which were similar to 0.01 and 0.16 reported by Berry et al. [4] for a New Zealand F1 Holstein x Jersey population used to produce milk with high percentage of total solids.

3.2. Genetic diversity

The analyzed loci were 100% polymorphic, similar to the findings for Criollo Colombiano, Brahman, Holstein, Braunvieh, and Criollo Lechero Tropical populations [32]. These results suggest that the loci frequencies remain without change regardless of breed or origin of the population.

The most polymorphic gene was $\beta\text{-casein}$ with five alleles. The following more polymorphic genes were $\kappa\text{-casein}$ and $\beta\text{-lactoglobulin}$

Table 3 Allelic and genotypic frequencies for β -lactoglobulin gene in the Mexican Jersey cattle populations.

	n	Allele	frequenc	ries	Genot	Genotype frequencies					
		A	В	С	AA	AB	BB	ВС	CC		
Cows	401	0.28	0.71	0.01	0.10	0.37	0.51	0.02	N/P		
Sires	52	0.13	0.82	0.05	0.04	0.18	0.72	0.04	0.02		
Total	453	0.26	0.72	0.02	0.09	0.34	0.54	0.02	0.01		

Table 4Estimated values of observed heterozygosity (H_o) and expected heterozygosity (H_e) for the loci CSN2. CSN3. and LGB in the Mexican lersey population.

Locus	Но	Не	X ²
CSN2	0.60	0.46	0.033 ^a
CSN3	0.50	0.45	
LGB	0.50	0.41	
Population	0.53	0.44	

^a X² test *P* value (*P* value < 0.05, not consistent with Hardy Weinberg Equilibrium).

with three observed alleles each one. The three genes had one effective allele. This is, when comparison for the average observed alleles to the average effective alleles through the three loci was done, the number of effective alleles was 30% less than those observed.

3.3. Heterozygosity and Hardy-Weinberg equilibrium

The studied population showed up differences (P < 0.05) between observed (H_o) and expected (H_e) heterozygosity (Table 4). These findings indicate that H_o doubled H_e in populations where reproductive management and genetic improvement programs have been applied. Similar results were reported by other authors in Holstein, Brahman, Braunvieh and Jersey populations [11,31]. In those populations matings were planned, therefore some influence from artificial selection for an economic trait could be expected.

4. Conclusion

The sampled Mexican Jersey population has a diverse genetic pool. The most common alleles were A^2 for β -casein, and B for both κ -casein and β -lactoglobulin. The Mexican Jersey population has the potential to be used in genetic improvement programs, aimed to improve the milk quality traits of economic importance.

Financial support

This project was supported by CONARGEN (2013-001) (Mexican Council of Livestock Genetic Resources) and Universidad Autónoma Chapingo, Mexico (DGIP-11550301). The authors also thank CONACYT for the financial support to the first author during his Master of Science studies.

References

- [1] Larios-Sarabia N, Ramírez-Valverde R, Núñez-Domínguez R, García-Muñiz JG, Ruíz-Flores A. Caracterización técnica, social y económica de las empresas del hato bovino jersey de registro en México. Agricultura, Sociedad y Desarrollo 2011;8:229-47.
- [2] Van Eenennaam AL, Medrano JF. Milk protein polymorphisms in California dairy cattle. J Dairy Sci 1991;74:1730–42. http://dx.doi.org/10.3168/jds.S0022-0302(91)78336-7.
- [3] Wedholm A, Larsen LB, Lindmark-Mansson H, Karlsson AH, Andrén A. Effect of protein composition on the cheese-making properties of milk from individual dairy cows. J Dairy Sci 2006;89:3296-305. http://dx.doi.org/10.3168/jds.S0022-0302(06)72366-9.
- [4] Berry SD, López-Villalobos N, Beattie EM, Davis SR, Adams LF, Thomas NL, et al. Mapping a quantitative trait locus for the concentration of β-lactoglobulin in milk, and the effect of β-lactoglobulin genetic variants on the composition of milk from Holstein-Friesian x Jersey crossbred cows. N Z Vet J 2010;58:1–5. http://dx.doi.org/10.1080/00480169.2010.65053.
- [5] Gibson JP, Jansen GB, Rozzi P. The use of κ-casein genotypes in dairy cattle breeding. Proceedings of the 4th World congress on genetics applied to livestock production July 23rd to 27th. Edinburgh, Scotland; 1990. p. 163–6.
- [6] Jo udo I, Henno M, Kaart T, Pussa T, Kart O. The effect of milk protein contents on the rennet coagulation properties of milk from individual dairy cows. Int Dairy J 2008; 18:964–7. http://dx.doi.org/10.1016/j.idairyj.2008.02.002.
- [7] Villa-Angulo R, Matukumalli LK, Gill CA, Choi J, Van Tassell CP, Grefenstette JJ. High-resolution haplotype block structure in the cattle genome. BMC Genet 2009; 10:19. http://dx.doi.org/10.1186/1471-2156-10-19.
- [8] Van Raden PM, Olson KM, Wiggans GR, Cole JB, Tooker ME. Genomic inbreeding and relationships among Holsteins, Jerseys and Brown Swiss. J Dairy Sci 2011;94: 5673–82. http://dx.doi.org/10.3168/jds. 2011-4500.

- [9] Medrano JF, Aguilar-Cordova E. Polymerase chain reaction amplification of bovine β-lactoglobulin genomic sequences and identification of genetic variants by RFLP analysis. Anim Biotechnol 1990;1:73–7. http://dx.doi.org/10.1080/10495399009525730.
- [10] Mitra A, Schlee P, Krause I, Blusch J, Werner T, Balakrishnan CR, et al. Kappa-casein polymorphisms in Indian dairy cattle and Buffalo: A new genetic variant in Buffalo. Anim Biotechnol 1998;9:81–7. http://dx.doi.org/10.1080/10495399809525896.
- [11] Ren DX, Miao SY, Chen YL, Zou CX, Liang XW, Liu JX. Genotyping of the k-casein and β-lactoglobulin genes in Chinese Holstein, Jersey and water Buffalo by PCR-RFLP. J Genet 2011:90:1–5. http://dx.doi.org/10.1007/s12041-011-0048-z.
- [12] NCBI (National Center of Biotechnology Information). GenBank: SNP database. [cited 31 March 2012]. Available from Internet: http://www.ncbi.nlm.nih.gov/snp.
- [13] Farrell Jr HM, Jimenez-Flores R, Bleck GT, Brown EM, Butler JE, Creamer LK, et al. Nomenclature of the proteins of cows' milk — Sixth revision. J Dairy Sci 2004;87: 1641–74. http://dx.doi.org/10.3168/jds.S0022-0302(04)73319-6.
- [14] Yeh CF, Yang RC, Boyle T. PopGen32 program. Canada: University of Alberta; 1999.
- [15] Bech AM, Kristiansen KR. Milk protein polymorphism in Danish dairy cattle and the influence of genetic variants on milk yield. J Dairy Res 1990;57:53–62. http://dx.doi.org/10.1017/S0022029900026601.
- [16] Winkelman AM, Wickham BW. Associations between milk protein genetic variants and production traits in New Zealand dairy cattle. Milk protein polymorphism. Proc. IDF Seminar held in Palmerston North, New Zealand. Int Dairy Fed; 1997. p. 38-46.
- [17] Miciński J, Klupczyński J, Mordas W, Zabłotna R. Yield and composition of milk from Jersey cows as dependent on the genetic variants of milk proteins. Pol J Food Nutr Sci 2007;57:95–9.
- [18] Tailford KA, Berry CL, Thomas AC, Campbell JH. A casein variant in cow's milk is atherogenic. Atherosclerosis 2003;170:13–9. http://dx.doi.org/10.1016/S0021-9150(03)00131-X.
- [19] Kami nski S, Cie sli nska A, Kostyra E. Polymorphism of bovine beta-casein and its potential effect on human health. J Appl Gen 2007;48:189–98. ettam GR, Adams LB. How to prepare an electronic version of your article. In: Jones BS, Smith RZ, editors. Introduction to the electronic age. New York: E-Publishing Inc.; 1999. p. 281–304.
- [20] Swaisgood HE. Chemistry of the caseins. In: Fox PF, editor. Advanced dairy chemistry I: Proteins. 2nd ed. New York, United States of America: Elsevier Applied Science; 1992. p. 63–110.
- [21] Miluchová M, Trakovická A, Gábor M. Analysis of polymorphism of beta casein of Slovak Pinzgau cattle by PCR-RFLP for alleles A1 and A2. Zootehnie şi Biotehnologii 2009;42:288–92.

- [22] Gurcan EK. Association between milk protein polymorphism and milk production traits in black and white dairy cattle in Turkey. African. J Biotechnol 2011;10: 1044–8.
- [23] Ng-Kwai-Hang KF, Grosclaude F. Genetic polymorphism of milk proteins. Adv Dairy Chem 2003;1:739–816. http://dx.doi.org/10.1007/978-1-4419-8602-3_22.
- [24] Ehrmann S, Bartenschlager H, Geldermann H. Quantification of gene effects on single milk proteins in selected groups of dairy cows. J Anim Breed Genet 1997;114: 121–32. http://dx.doi.org/10.1111/j.1439-0388.1997.tb00499.x.
- [25] Çardak AD. Effects of genetic variants in milk protein on yield and composition of milk from Holstein-Friesian and Simmentaler cows. S Afr J Agric Sci 2005;35:41–7.
- 26] Hanusová E, Huba J, Oravcová M, Polák P, Vrtková I. Genetic variants of beta casein in Holstein dairy cattle in Slovakia. Slovak J Anim Sci 2010;43:63–6.
- [27] Trujillo E, Noriega D, Camargo M. Genotipificación de kappa-caseína bovina y evaluación de las frecuencias genotípicas y alélicas de sus polimorfismos en cuatro razas. Actual Biol 2000:22:145–52.
- [28] Jann OC, Ibeagha-Awemu EM, Özbeyaz C, Zaragoza P, Williams JL, Ajmone-Marsan P, et al. Geographic distribution of haplotype diversity at the bovine casein locus. Genet Sel Evol 2004;36:243–57. http://dx.doi.org/10.1186/1297-9686-36-2-243.
- [29] Tsiaras AM, Bargouli GG, Banos G, Boscos CM. Effect of kappa-casein and beta-lactoglobulin loci on milk production traits and reproductive performance of Holstein cows. J Dairy Sci 2005;88:327–34. http://dx.doi.org/10.3168/jds.S0022-0302(05)72692-8.
- [30] Boetcher PJ, Caroli A, Stella A, Chessa S, Budelli E, Canavesi F, et al. Effects of casein haplotypes on milk production traits in Italian Holstein and Brown Swiss cattle. J Dairy Sci 2004;87:4311–7. http://dx.doi.org/10.3168/jds.S0022-0302(04)73576-6.
- [31] Hernández A, Salinas VM, Luna M, García R, Cervantes P, Uffo O, et al. Comportamiento de las variantes genéticas de los genes k-caseína, alfa-lactoalbúmina y hormona del crecimiento en la raza criollo lechero tropical en México y su relación con el nivel de respuesta al calor. Rev Salud Anim 2006;28:96–104.
- [32] Cervantes P, Luna M, Hernández A, Pérez-Gil F, Ponce P, Uffo O. Polimorfismo genético en el locus de la kappa-caseína, en vacas de diferentes razas y cruces en el trópico mexicano. Rev Salud Anim 2007;29:78–84.
- 33] Meza-Nieto MA, González-Córdova AF, Becerril-Pérez CM, Ruíz-López FJ, Díaz-Rivera P, Vallejo-Córdoba B. Polimorfismo genético de la β-lactoglobulina en la leche de vacas Holstein y Criollo Lechero Tropical. Agrociencia 2010;44:531–9.