

Draft genome sequences of three NDM-1-producing Enterobacteriaceae species isolated from Brazil

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The emergence of multidrug-resistant Enterobacteriaceae strains producing carbapenemases, such as NDM-1, has become a major public health issue due to a high dissemination capacity and limited treatment options. Here we describe the draft genome of three NDM-1-producing isolates: Providencia rettgeri (CCBH11880), Enterobacter hormaechei subsp. oharae (CCBH10892) and Klebsiella pneumoniae (CCBH13327), isolated in Brazil. Besides bla_{NDM-1} resistance genes to aminoglycosides [aadA1, aadA2, aac(6')-Ib-cr] and quinolones (qnrA1, qnrB4) were observed which contributed to the multidrug resistance profile. The element ISAba125 was found associated to the bla_{NDM-1} gene in all strains.

Key words: NDM-1 - Brazil - Enterobacteriaceae

The emergence of carbapenemase-producing Enterobacteriaceae has become a major public health issue worldwide due to a high dissemination capacity and limited treatment options (Nordaman et al. 2011). NDM is a metallo-beta-lactamase first reported in 2009 (Yong et al. 2009) and, now, it has already been detected in several countries worldwide. In Brazil, this carbapenemase was first described in a *Providencia rettgeri* isolate from the city of Porto Alegre, state of Rio Grande do Sul (RS) (South Region of Brazil), in 2013 (Carvalho-Assef et al. 2013). Then, the detection of six clonally related NDM-producing *Enterobacter hormaechei* subsp. *oharae* isolates was reported from the same public hospital (Carvalho-Assef et al. 2014). After that, nine NDM-1-producing *Enterobacter cloacae* complex isolates and two *Morganella morganii* were also recovered from three different hospitals in Porto Alegre (Rozales et al. 2014). Recently, NDM-1-producing bacteria have been described in the state of Rio de Janeiro (RJ) (Southeast Region of Brazil) (Pereira et al. 2014). In 2014, a NDM-1-producing *Acinetobacter baumannii* was also detected in Londrina, state of Paraná (South Region of Brazil) (Pillonetto et al. 2014).

Here, we aim to announce the draft genome of three NDM-1-producing isolates: *P. rettgeri* (CCBH11880), isolated from a surgical wound of a patient from RS, *E. hormaechei* subsp. *oharae* (CCBH10892), isolated from a rectal swab of another patient in RS, and *Klebsiella pneumoniae* (CCBH13327), isolated from a rectal swab of a patient in the city of Rio de Janeiro, RJ.

Initially all of the strains were tested against different antimicrobial drugs and showed multidrug resistance profiles. Genomic DNA of all strains was extracted using QIAamp DNA Blood Mini Kit (Qiagen, Germany). Whole genome shotgun libraries from each strain were prepared with the Nextera XT DNA Sample Prep kit (Illumina Inc, USA), according to the manufacturer's instructions, and sequenced on an Illumina MiSeq system with the MiSeq Reagent v.2 500 cycles kit. Sequence reads were then trimmed and filtered using a Phred score >20. The software Geneious v.6.1.7 (Biomatters Ltd, New Zealand) was used to perform *de novo* assembling. Rapid Annotation using System Technology v.2.0 server was used for genome annotation. Acquired resistance genes were analysed using the ResFinder platform (genomicpidemiology.org). The detailed features of all isolates can be found on Table.

For the *P. rettgeri* isolate (CCBH11880) we obtained 656,560 paired end reads of 250 base pairs (bp), which were assembled into 80 contigs. The G+C content for this strain was 41%, considered common for this species. The estimated genome size, comprising all contigs, was 4,999,177 bp. Overall, 4,670 protein coding sequences were found and 89 RNAs were annotated (79 tRNA and 10 rRNA). Acquired resistance genes were searched using the ResFinder platform and different resistance genes were observed such as: *aadA1* (GenBank JSEQ01000006.1; 199,767-200,555 bp), *strA* (GenBank JSEQ01000017.1; 91,398-92,201 bp), *strB* (GenBank JSEQ01000017.1; 92,201-93,037 bp), *aadB* (GenBank JSEQ01000041.1; 326-859 bp), *aac(6')-Ib* (GenBank JSEQ01000017.1; 2,882-3,400 bp), *qnrD* (GenBank JSEQ01000028.1; 1,671-2,315 bp), *bla*_{OXA-10} (GenBank JSEQ01000017.1; 2,014-2,814 bp), *bla*_{NDM-1} (GenBank JSEQ01000024.1; 412-1,224 bp), *ere(A)* (GenBank JSEQ01000025.1; 295-1,521 bp), *msr(E)* (GenBank JSEQ01000023.1; 37,090-38,565 bp), *mph(E)* (GenBank JSEQ01000023.1; 36,150-37,034 bp), *floR* (GenBank JSEQ01000017.1; 96,173-97,386 bp), *catA1* (GenBank JSEQ01000031.1; 870-1,529 bp), *sul1* (GenBank JSEQ01000017.1; 3,823-4,749 bp), *sul2* (GenBank

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TABLE
Genetic information about three NDM-1-producing isolates from Brazil

Isolate feature	<i>Providencia rettgerii</i> (CCBH11880)	<i>Enterobacter hormaechei</i> (CCBH10892)	<i>Klebsiella pneumoniae</i> (CCBH13327)
NCBI accession	JSEQ000000000	JSBO000000000	JSER000000000
BioProject	PRJNA264579	PRJNA264581	PRJNA264954
Isolation source	Surgical wound	Rectal swab	Rectal swab
City/state of origin	Porto Alegre/RS	Porto Alegre/RS	Rio de Janeiro/RJ
GC content (%)	41	54.5	56.6
Paired end reads (n)	656,560	2,283,589	1,748,579
Genome coverage	32X	106X	72X
Estimated genome size (bp)	4,999,177	5,373,562	6,023,847
Contigs (n)	80	58	106
N50	282.487	277.989	133.213
Coding sequences (n)	4,670	5,134	5,722
RNAs (n)	89	102	99
tRNA (n)	79	89	86
rRNA (n)	10	13	13
Resistance genes	<i>aadA1</i> , <i>strA</i> , <i>strB</i> , <i>aadB</i> , <i>aac(6')-Ib</i> , <i>qnrD</i> , <i>bla</i> _{OXA-10} , <i>bla</i> _{NDM-1} , <i>ere(A)</i> , <i>msr(E)</i> , <i>mph(E)</i> , <i>floR</i> , <i>catA1</i> , <i>sulI</i> , <i>sul2</i> , <i>tet(A)</i> , <i>dfrA1</i> , <i>dfrA8</i>	<i>aadA2</i> , <i>aph(3')-Ia</i> , <i>strA</i> , <i>strB</i> , <i>bla</i> _{DHA-1} , <i>bla</i> _{ACT-7} , <i>bla</i> _{CTX-M-15} , <i>bla</i> _{NDM-1} , <i>qnrB4</i> , <i>ere(A)</i> , <i>sulI</i> , <i>tet(A)</i> , <i>tet(D)</i>	<i>aadA2</i> , <i>aac(3)-IIa</i> , <i>bla</i> _{SHV-99} , <i>bla</i> _{CTX-M-2} , <i>bla</i> _{CARB-2} , <i>bla</i> _{NDM-1} , <i>oqxA</i> , <i>oqxB</i> , <i>qnrA1</i> , <i>catA1</i> , <i>sulI</i> , <i>tet(D)</i> , <i>dfrA8</i>

bp: base pair; NCBI: National Center for Biotechnology Information; RJ: state of Rio de Janeiro; RS: state of Rio Grande do Sul.

JSEQ01000017.1; 90,522-91,337 bp), *tet(A)* (GenBank JSEQ01000017.1; 94,373-95,544 bp), *dfrA1* (GenBank JSEQ01000006.1; 198,617-199,090 bp) and *dfrA8* (GenBank JSEQ01000033.1; 334-843 bp).

Genome sequencing of *E. hormaechei* subsp. *oharae* isolate (CCBH10892) generated 2,283,589 paired end reads of 250 bp, yielding 58 contigs after assembly (Geneious v.6.1.7) for a genome size estimative of 5,373,562 bp. A total of 5,134 protein coding sequences and 102 RNAs were observed, being 89 tRNA and 13 rRNA. ResFinder analysis provided the following acquired resistance genes: *aadA2* (GenBank JSBO01000022.1; 2,498-3,289 bp), *aph(3')-Ia* (GenBank JSBO01000057.1; 354-932 bp), *strA* (GenBank JSBO01000042.1; 2,178-2,981 bp), *strB* (GenBank JSBO01000042.1; 2,981-3,817 bp), *bla*_{DHA-1} (GenBank JSBO01000036.1; 13,727-14,866 bp), *bla*_{ACT-7} (GenBank JSBO01000001.1; 594,635-595,780 bp), *bla*_{CTX-M-15} (GenBank JSBO01000007.1; 210,876-211,751 bp), *bla*_{NDM-1} (GenBank JSBO01000041.1; 4,501-5,313 bp), *qnrB4* (GenBank JSBO01000036.1; 8,959-9,606 bp), *ere(A)* (GenBank JSBO01000002.1; 405,895-406,937 bp), *sulI* (GenBank JSBO01000022.1; 3,707-4,633 bp), *tet(D)* (GenBank JSBO01000046.1; 2,098-3,297 bp) and *tet(A)* (GenBank JSBO01000044.1; 1,151-2,335 bp).

The *K. pneumoniae* isolate (CCB13327) had an estimated genome size of 6,023,847 bp. An assembly (Geneious v.6.1.7) with 106 contigs was achieved with 1,748,579 paired end reads of 250 bp. A total of 5,722 protein coding sequences were observed, including 99 RNA sequences (86 tRNA and 13 rRNA). Ac-

quired resistant genes found were: *aadA2* (GenBank JSER01000014.1; 552-1,343 bp), *aac(3)-IIa* (GenBank JSER01000058.1; 14,738-15,598 bp), *bla*_{SHV-99} (GenBank JSER01000008.1; 101,044-101,904 bp), *bla*_{CTX-M-2} (GenBank JSER01000045.1; 2,516-3,391 bp), *bla*_{CARB-2} (GenBank JSER01000014.1; 1,461-2,375 bp), *bla*_{NDM-1} (GenBank JSER01000063.1; 4,749-5,561 bp), *oqxA* (GenBank JSER01000088.1; 4,991-6,166 bp), *oqxB* (GenBank JSER01000088.1; 2,518-4,967 bp), *qnrA1* (GenBank JSER01000080.1; 416-1,072 bp), *catA1* (GenBank JSER01000041.1; 3,303-3,962 bp), *sulI* (GenBank JSER01000058.1; 4,497-5,423 bp), *tet(D)* (GenBank JSER01000091.1; 392-1,576 bp) and *dfrA8* (GenBank JSER01000058.1; 816-1,325 bp). Overall, we observed that these NDM-1-producing Enterobacteriaceae strains carry resistance genes to different antimicrobial classes, which can explain the multidrug resistant profile observed.

In *P. rettgeri* CCBH 11880, the *bla*_{NDM-1} was found inside the Tn125 transposon, which is composed of two copies of IS*Aba125* (Carvalho-Assef et al. 2013) (data not shown). In the *E. hormaechei* (Carvalho-Assef et al. 2014) (GenBank accession NG041719) and *K. pneumoniae* isolates, the *bla*_{NDM-1} gene was found flanked by only one copy of IS*Aba125* (truncated) at the right boundary (data not shown).

The announcement of the whole-genome sequence of the three NDM-1-producing Enterobacteriaceae strains also provides basis for other studies, which will certainly increase our understanding of the role of this species in the drug resistance scenario.

Nucleotide sequence accessions - These Whole Genome Shotgun project have been deposited in DDBJ/ENA/GenBank under the accessions JSEQ000000000, JSBO000000000 and JSER000000000 for *P. rettgerii* (CCBH11880), *E. hormaechei* (CCBH10892) and *K. pneumoniae* (CCBH13327), respectively. The versions described in this paper are the first version (JSEQ000000000, JSBO000000000 and JSER000000000).

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