

Primary dengue haemorrhagic fever in patients from northeast of Brazil is associated with high levels of interferon- β during acute phase

Renato Antônio dos Santos Oliveira¹, Mayara Marques Carneiro da Silva¹,
Carlos Eduardo Calzavara-Silva², Ana Maria Silva¹, Marli Tenório Cordeiro¹,
Patrícia Muniz Mendes Freire de Moura⁴, Paulo Neves Baptista Filho⁵,
Ernesto Torres de Azevedo Marques Júnior^{1,3}, Laura Helena Vega Gonzales Gil^{1/+}

¹Fundação Oswaldo Cruz, Centro de Pesquisas Aggeu Magalhães, Departamento de Virologia, Recife, PE, Brasil

²Fundação Oswaldo Cruz, Centro de Pesquisas René Rachou, Departamento de Imunologia, Laboratório de Imunologia Celular e Molecular, Belo Horizonte, MG, Brasil ³University of Pittsburgh, Center for Vaccine Research, Department of Infectious Diseases and Microbiology, Pittsburgh, PA, USA ⁴Universidade de Pernambuco, Instituto de Ciências Biológicas, Recife, PE, Brasil ⁵Universidade de Pernambuco, Faculdade de Ciências Médicas, Recife, PE, Brasil

Dengue is an acute febrile disease caused by the mosquito-borne dengue virus (DENV) that according to clinical manifestations can be classified as asymptomatic, mild or severe dengue. Severe dengue cases have been associated with an unbalanced immune response characterised by an over secretion of inflammatory cytokines. In the present study we measured type I interferon (IFN-I) transcript and circulating levels in primary and secondary DENV infected patients. We observed that dengue fever (DF) and dengue haemorrhagic fever (DHF) patients express IFN-I differently. While DF and DHF patients express interferon- α similarly ($52,71 \pm 7,40$ and $49,05 \pm 7,70$, respectively), high levels of circulating IFN- β were associated with primary DHF patients. On the other hand, secondary DHF patients were not able to secrete large amounts of IFN- β which in turn may have influenced the high-level of viraemia. Our results suggest that, in patients from our cohort, infection by DENV serotype 3 elicits an innate response characterised by higher levels of IFN- β in the DHF patients with primary infection, which could contribute to control infection evidenced by the low-level of viraemia in these patients. The present findings may contribute to shed light in the role of innate immune response in dengue pathogenesis.

Key words: dengue fever - dengue haemorrhagic fever - type I interferon - innate immunity

Dengue is an acute febrile disease caused by a small single-stranded RNA virus with four antigenically distinct serotypes [dengue virus (DENV)-1 to 4]. DENV belongs to the genus *Flavivirus* of the *Flaviviridae* family and is transmitted to humans by infected *Aedes aegypti* mosquitoes (Ross 2010). DENV is circulating in more than 100 countries worldwide and causes 400 million new cases each year, of which only 100 million are symptomatic (Messina et al. 2015). DENV infection presents different clinical manifestations ranging from asymptomatic to severe. Symptomatic DENV infection is classified according to the severity of the disease in dengue fever (DF) or dengue haemorrhagic fever (DHF). DF is a self-limiting febrile illness, whereas DHF is a life-threatening condition characterised by an increased capillary permeability, thrombocytopenia and bleeding events that could lead to shock (Back & Lundkvist 2013).

Alternatively, the WHO proposed a new classification system based on clinical and laboratorial parameters. This classification stratifies the dengue cases in dengue without warning signs, dengue with warning signs and severe dengue (WHO 2009). Different theories have tried to explain the pathogenesis of dengue infection (Martina et al. 2009), however the precise mechanism that triggers DHF remains poorly understood, this way an early identification of factors that could predispose the development of DHF would be of great clinical relevance. One of the hypotheses, the antibody-dependent enhancement (ADE), proposes that DHF could be a consequence of an unbalanced immune response mediated by non-neutralising antibodies (Halstead & O'Rourke 1977, Halstead 1988). This condition would be characterised by increased monocyte activation and secretion of chemical mediators and pro-inflammatory cytokines (Green & Rothman 2006, Kurane 2007). Differences in the cytokines (TNF- α , IFN- γ , IL-6 IL-13) (Dong et al. 2007, Priyadarshini et al. 2010) and chemokines (IL-8, MIP, IP-10, MCP-1, MIP-1 β) production (Chen et al. 2006, Lee et al. 2006, Fink et al. 2007, Assunção-Miranda et al. 2010), in culture supernatants and plasma samples from DF and DHF patients have been described. High serum levels of the pro-inflammatory cytokines IFN- γ and TNF- α have been associated with disease severity (Nguyen et al. 2004), while Macrophage Inflammatory Protein-1 β (MIP-1 β) has been associated with a good prognosis (Bozza et al. 2008).

doi: 10.1590/0074-02760150453

Financial support: NIAID/NIH (grants U19 AI56541 and N01 AI 40085), CNPq (grants CNPq-550140/2010-7 and CNPq-404453/2012-0 (RASO's visiting research fellowship)), FACEPE (grant APQ-1516-4.01/10 (MTC's research fellowship)).

RASO and MMCS contributed equally to this work.

+ Corresponding author: laura@cpqam.fiocruz.br

Received 14 December 2015

Accepted 29 April 2016

Mechanisms of innate immune response mediated by interferon (IFN) represent the most important pathways of host defense directed to hinder viral replication. The name interferon arose precisely because of its ability to interfere with viral replication. The IFN family consists of IFNs type I, II and III. Type I interferons (IFN-I) include 13 different IFN- α subtypes and one each of IFN- β , IFN- ω , IFN- ϵ and IFN- κ , they are the most important cytokines involved in the control of viral infection and can be secreted by most nucleated cells (Sen 2001, Sadler & Williams 2008), specially by plasmacytoid dendritic cells (Siegal et al. 1999). The type I IFN secreted by virus infected cells acts in an paracrine and autocrine manner, through the binding to surface receptors initiating a signaling cascade through the JAK/STAT pathway which ends up regulating the expression of interferon-induced genes (ISG). These genes encode proteins that promote an “antiviral” state in both infected and non-infected cells, which inhibits the virus life cycle, hampering its spreading (Samuel 2001). In order to become successful pathogens, viruses have developed different strategies to subvert the antiviral activity of type I IFN. Many viruses encode proteins that antagonise key steps of either type I IFN induction or signaling pathways (Jones et al. 2005, Umareddy et al. 2008, Ashour et al. 2009, Li et al. 2013, Wu et al. 2013). The importance of type I IFN for DENV control was evidenced by the IFN- β dependence of cultured hepatocytes to control virus replication (Liang et al. 2011).

Tang et al. (2010) described IFN- α and IFN- β levels in a cohort of hospitalised DENV-1 patients and IFN- α levels were more elevated than IFN- β which levels were similar to uninfected control (Tang et al. 2010). Maximum plasma levels of IFN- α were generally not seen and took place before to peak viraemia in DENV-3 infected children (Libraty et al. 2002), and more recently in an elegant study, Gandini et al. 2013 demonstrated that circulating plasma levels of IFN- α were higher in mild dengue compared with severe dengue or healthy controls (Gandini et al. 2013). Although the levels of IFN- α has been well described, little is known about IFN- β circulating levels during the acute phase of infection. This report describes for the first time differences of circulating IFN- β between DF and DHF patients during the acute phase of disease, up to five days after onset of symptoms. The differences on IFN- β levels observed could lead to altered immune response which may play an important role on dengue pathogenesis.

MATERIALS AND METHODS

Patients and samples - Blood samples were obtained from a cohort of 104 acute febrile dengue patients admitted to different hospitals in the city of Recife (Pernambuco, Brazil), as already described (Cordeiro et al. 2007a, b). Briefly, blood samples were sequentially obtained from each patient on the day of admission (day 1) and at days 3, 5, 7, 15 and 30. Dengue cases were confirmed by DENV isolation and/or viral RNA detection by reverse transcriptase polymerase chain reaction (RT-PCR) and/or by positive serology for anti-dengue IgM ELISA. Subsequently, samples were clinically classified

as DF or DHF according to 1997 WHO criteria (WHO 1997). Data about demographic characteristics, clinical features and biochemical and haematological laboratory tests were also collected while patients were hospitalised. Cytokine assays were carried out on 44 serum samples from DF patients and 33 serum samples from DHF patients. The presence of IgM and/or IgG antibodies specific to dengue virus was detected using a capture ELISA Kit, according to the manufacturer's instructions (E-DEN01M and E-DEN01G, PanBio).

Interferon mRNA levels by real-time RT-PCR - Quantitative real-time PCR (qPCR) assays were performed with peripheral blood mononuclear cells (PBMC) samples collected and purified from clinically classified DHF and DF patients by density gradient technique (Ficoll-Paque™ Plus- GE Healthcare, Uppsala-SE). At different time point of infection as follows: (i) During the acute phase, up to five days after the onset of symptoms; (ii) During the non-viraemic but symptomatic phase, from six-10 days after the onset of symptoms; and (iii) During the convalescent phase, > 10 days after the onset of symptoms.

Genes were amplified and detected using TaqMan® gene expression assays (Applied BioSystems, cat. 4331182 - gene id: IFN α -1: Hs00855471_g1; IFN β -1: Hs01077958_s1 and IFN γ : Hs00989291_m1). Total RNA was extracted using the RNeasy Mini Kit (Qiagen) and treated with DNase (Qiagen) following the manufacturer's protocols. Total RNA (1 μ g) was reverse transcribed to cDNA using a Super-Script III First-Strand Synthesis System (Invitrogen) and Random Hexamer Primers (Invitrogen) under the following reaction conditions: 50°C for 30 min, 85°C for 5 min and then incubation on ice. RNase H (2 U) (Invitrogen) was added and samples were incubated at 37°C for 20 min. qPCR was performed using the ABI PRISM 7500 (Applied BioSystems). cDNA obtained from the total RNA from the patients described above was used. β -actin gene was used to normalise the gene data. Reactions were performed in triplicate and contained 2 μ L of cDNA, 6.25 μ M of each specific primer (Applied BioSystems), TaqMan Universal PCR Master Mix (Applied BioSystems) and water, in a final volume of 25 μ L. Triplicates of non-template controls were included for each qPCR experiment. Cycle conditions were as follows: after initial periods of 2 min at 50°C and 10 min at 95°C, samples were cycled 40 times at 95°C for 15 s and 60°C for 1 min. The baseline and threshold for cycle threshold (Ct) calculations were set automatically using Sequence Detection Software, version 1.4 (Applied BioSystems). The efficiency of amplification (E) of each target molecule was calculated from the slope of the standard curve (plot of Ct vs. the negative log10 concentration of the target) derived from the slopes {E = [10(-1/Slope)]-1}. Once all assays met the amplification efficiency criteria of 100% \pm 10% (Livak & Schmittgen 2001), the 2- $\Delta\Delta$ Ct method was used for relative calculations (Applied Biosystems 2010, 2012, Livak & Schmittgen 2001).

Measurements of type I IFN levels by ELISA - The levels of IFN-I (α & β) in the serum/plasma of patients up to five days after onset of symptoms were evaluated by quantitative enzyme-linked immunosorbent assay

(ELISA). The lower limits of detection were 1.0 pg/mL and 2.3 pg/mL for the human IFN- α (My Biosource, San Diego, CA) and IFN- β (Axxora, Farmingdale, NY) kits, respectively. Optical densities at 450nm were measured on an ELISA dedicated instrument and cytokine concentrations were obtained using a standard curve. All determinations were performed in duplicates and arithmetic averages were calculated.

Real time RT-PCR assay for dengue - Viraemia was quantified and expressed as the number of RNA copies per mL. Viral RNA was extracted from 140mL of each individual serum sample using the QIAamp Viral RNA Mini Kit/QIAamp MiniElute Virus Spin Reagent (Qiagen, Valencia, CA). Reverse transcriptions were performed using random primers (hexamers), RNase inhibitors (RNase OUT 40 U/mL), Superscript III reverse transcriptase 200 U/mL and 5mL of purified RNA, according to the manufacturer's specifications (Invitrogen). Quantification was performed by Real Time PCR with SYBR Green-based specific to DENV3 serotype kit for dengue (DSSS-P29), kindly provided by Dr Hen Phon-Too (National University of Singapore), able to detect down to a minimum of 10 RNA copies/mL. Each RT-PCR reaction included 32 mL of the DSSS kit mix with primers (0.2 mM), Xtensa[®] PCR buffer (Bioworks, Singapore), MgCl₂ (2.5 mM) and 0.5 mL Platinum hot start polymerase (5U/mL; Invitrogen), 2 mL reverse transcription mix and water, making a final volume of 50 mL. Primers were synthesised with NS4A sequence tags (forward primers with 5'GTCAGAA(C/G)ATG-GCGGTAGG3' and reverse primers with 5' CTTTC-CAATCCCTTTACCTGATAT3'). Amplification conditions were as follows: 2 min at 50°C, 3 min at 95°C, then 40 cycles of 30 s at 95°C and 1 min at 60°C. qPCR assays were performed in the ABI PRISM 7500 Sequence Detection System (Applied Biosystems, CA, USA). ABI PRISM software (version 1.4) was used to analyse qPCR products considering the melting temperature (dissociation curve), amplification curve and standard deviation between duplicates for quantification. The efficiency of amplification was calculated for each plate using the threshold cycle value (Ct) and the slope of the standard curve. Standard curves were obtained by amplification of the appropriate standards (10⁷ to 10¹⁰ copies). False positive results due to contaminations were ruled out by running no-template controls (NTC). Sera from health humans were used as negative controls. The local DENV-3 isolate propagated in *Aedes albopictus* C6/36 cells (10⁶ DENV RNA copies) was used as positive control. All assays were performed in duplicates.

Statistical analysis - Statistical analyses were performed using GraphPad Prism version 4.0a for Macintosh OS X (GraphPad Software, San Diego, CA). Real-time PCR data were analysed with multiple comparison Kruskal-Wallis test with Dunn's correction. Fisher's exact test was used to evaluate frequencies of positivity in symptoms. Differences in demographic information between DF and DHF patients were evaluated by nonparametric Mann-Whitney U test. Cytokines levels were analysed by one-way analysis of variance (ANOVA) for mean compari-

son for multiple groups with post hoc Bonferroni's test, p values ≤ 0.05 were considered significant.

Ethics - The procedure followed was in accordance with the ethical standards of the committee on human experimentation of the Aggeu Magalhães Research Center as well as with Helsinki Declaration of 1975. A written informed consent was obtained from all subjects, and the protocol for this study was approved by Aggeu Magalhães Research Center committee under the number CEP: 11/11.

RESULTS

Clinical characteristics of DENV-infected patients - 104 patients (46% men and 54% women) with acute dengue infection were included in the present study, of which 48 (46%) were classified as DF and 56 (54%) as DHF. The characteristics of studied population are summarised in Table. Age and gender of DF and DHF groups were similar with no significant difference between their medians.

At acute phase of infection, leukopenia was a condition detected in both DF and DHF patients, 21 (47.7%) DHF patients and 16 (33.3%) DF patients presented leukopenia. Other clinical signs of dengue, such as abdominal pain, hypotension, and bleeding manifestations such as gingival bleeding, had been reported by 60.3%, 7.7%, and 23% of the patients, respectively. Bleeding can be associated with the reduction in platelets numbers, in fact 79% of patients with gingival bleeding were thrombocytopenic.

Once liver function can be unsettled by dengue infection, liver function tests such as total albumin, aspartate transaminase (AST), and alanine transaminase (ALT) were accessed for some patients. 19% of dengue infected patients presented low total albumin levels (< 3.5 g/dL), 70.3% high levels of AST (> 40 IU/L) and 52.6% high levels of ALT (> 55 IU/L). The average levels of total albumin, AST, and ALT were statistically different when compared DHF to DF values (Table).

Previous incidence of DENV infections was investigated and evidenced by the presence of circulating anti-DENV IgG, secondary infections were observed in 37 (77%) of DF and 41 (73%) of DHF patients (Table). DENV serotypes were identified by a serotype-specific RT-PCR assay, and we could observe that DENV-3 was the predominant DENV serotype infecting 27 patients, five patients were infected by DENV-2 and other three had DENV-1 infection.

Immunological features of patients with dengue infection - IFN mRNA levels - Aiming to verify the influence of DENV infection on its primary cellular targets, we conducted qPCR assays to measure the levels of the genes encoding the IFN α , β and γ proteins, during the acute and convalescent phase of DENV infection. We obtained cDNA from total RNA extracted from PBMC samples of DF and DHF as templates for qPCR. In Fig. 1 it is possible to observe that the levels of IFN γ are not significantly altered by DENV infection even after 10 or more days post-infection. Patients with the severe form of DHF, with up to five days of fever, presented higher levels of IFN-I (α/β) when compared to patients suffering from mild illness (DF).

TABLE
Demography, clinical and laboratorial characteristics of study cohort

Cohort characteristics ^a	Dengue fever		Dengue haemorrhagic fever		p
	Median (95% CI) ^{b†}	N	Median (95% CI)	N	
Age (years)	12 (16-25)	48	24.5 (22-32)	56	ns
Gender (M:F)		27:21		21:35	ns
IgG, + : -		37:11		41:15	ns
White Blood Cells (WBC) (10 ³ /mm ³)	5.0 (4.6-6.3)	46	3.8 (3.8-6.1)	42	0.1954
Platelets (10 ³ /mm ³)	164.5 (148.6-217.5)	48	74.5 (69.7-117.6)	42	< 0.0001^c
Total albumin (g/L)	4.5 (4.2-4.6)	26	3.6 (3.4-4.0)	21	0.0001^c
Aspartate Transaminase (AST) (IU/L)	40.6 (51.2-124.2)	34	70 (58.7-157.3)	23	0.0293^c
Alanine Transaminase (ALT) (IU/L)	28.2 (25.62-101.3)	34	63 (54.9-119.7)	23	0.0047^c
Abdominal pain		30 (62.5%) ^d		31 (58.5%)	0.5501 ^f
Hypotension		1 (2%)		7 (12.5%)	0.0662 ^f
Bleeding manifestation ^e		6 (12.8%)		18 (32.1%)	0.0206^f

M: male; F: female; ns: not significant; ^astudy population with 104 patients; ^bCI, 95% confidence interval; ^cMann-Whitney test (p < 0.05 in bold, significant); ^dnumber (%) of positive patients; ^egingival bleeding; ^ffisher's exact test (p < 0.05 in bold, significant).

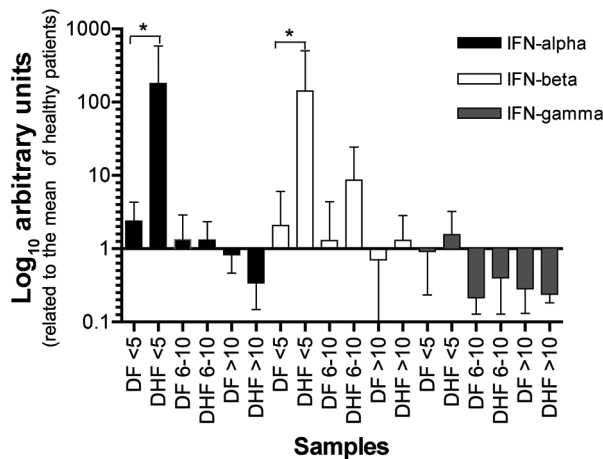


Fig. 1: quantitative real-time polymerase chain reaction (qRT-PCR) analysis of interferons (IFNs) α , β and γ transcript level in peripheral blood mononuclear cells (PBMC) obtained from dengue virus-infected patients. 59 patients [25 dengue haemorrhagic fever (DHF) patients, 29 dengue fever (DF) patients and five healthy volunteers] had their blood samples collected and used to extract PBMC and further to obtain total RNA and cDNA. Blood samples were collected in three phases of the disease: the acute phase (< 5 days post onset of symptoms), a putatively non-viraemic phase (between six-10 days post onset of symptoms) and the convalescent phase of disease (> 10 days post onset of symptoms). The average of mRNA type-I IFN of healthy donors was used as reference for baseline of IFNs expression and is represented as 1. Data were analysed with multiple comparison Kruskal-Wallis test with Dunn's correction. * = p < 0.05. DF n = 25 samples (< 5 days); n = 9 (six-10 days), and n = 5 (> 10 days). DHF n = 29 samples (< 5 days); n = 15 (six-10 days), and n = 15 (> 10 days).

Levels of circulating IFN- β - IFN-I plays an essential role in antiviral defenses (Sadler & Williams 2008). To determine whether high levels of IFN-I are associated to severe dengue cases during the acute phase, both IFN- β and IFN- α were measured in serum/plasma from DF and DHF patients within five days after the onset of symptoms. Fig. 2A shows that DENV patients presented similar levels of IFN- α (primary DF: 59.05 (11.4 ~ 201.2) pg/mL; secondary DF: 50.04 (0.08 ~ 221.2) pg/mL; primary DHF: 60.57 (1.75 ~ 165.3) pg/mL; secondary DHF: 43.30 (0 ~ 142.6) pg/mL). These values were not statistically different. Instead, primary DHF infected patients showed the highest levels of IFN- β (6.69 (0.15 ~ 34.9) pg/mL), and it is statistically higher than primary DF (1.57 (0 ~ 7.24) pg/mL), secondary DF (1.80 (0 ~ 9.77) pg/mL), and secondary DHF infected patients (2.35 (0 ~ 10.1) pg/mL) (Fig. 2B).

We tried to establish association with type I IFN levels and laboratorial biochemical parameters (AST, ALT, total albumin), as well as with haematological parameters (platelets and WBC). However, type I IFN expression was not statistically correlated with any biochemical or haematological parameter.

Viraemia levels in DF and DHF patients - The same serum samples used to determine IFN-I concentrations were also used to quantify DENV viraemia levels. Results showed that 29.5% and 24.2% of the samples were DENV-RNA positive in the DF and DHF groups, respectively. Fig. 3 shows the medians of viral load in DF and DHF patients divided in primary and secondary infections. The difference observed in the virus load between secondary DF (6.66 log₁₀ RNA copies/mL) and

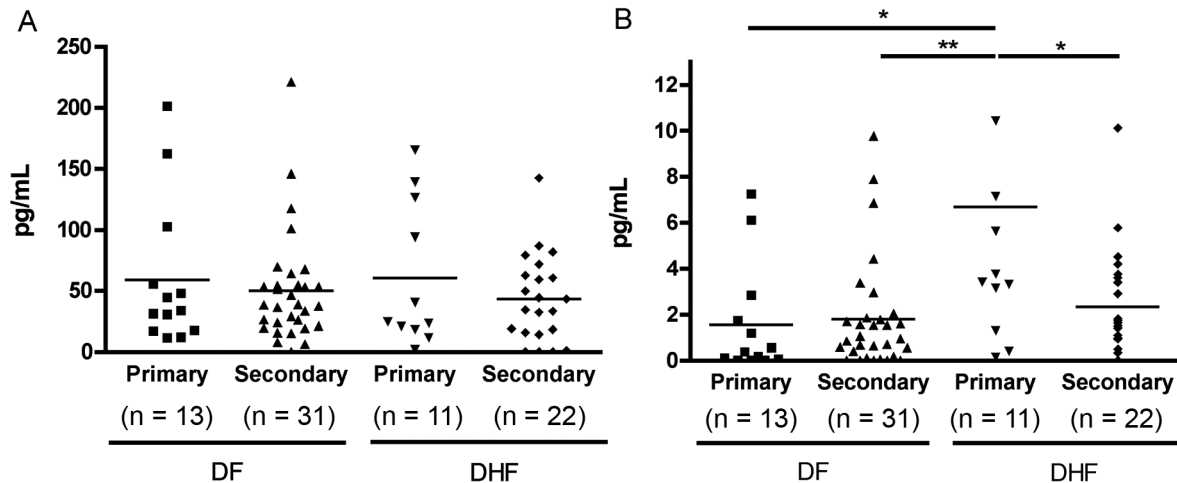


Fig. 2: different pattern of type I interferon (IFN) production in acute primary and secondary dengue virus infections. Serum levels of IFN- α (A) and IFN- β (B) from 44 dengue fever patients and 33 dengue haemorrhagic fever patients during acute phase of infection (< 5 days post onset of symptoms) are presented. Each dot represents one subject. All results are expressed as pg/mL. * = $p < 0.05$, and ** = $p < 0.01$ in Bonferroni's post hoc test.

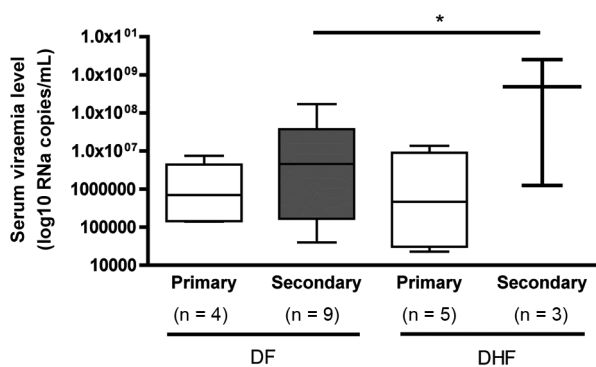


Fig. 3: viraemia levels in patients with primary and secondary dengue virus infections. Box-plot distribution of viraemia levels by primary and secondary dengue infection in dengue fever (DF) and dengue haemorrhagic fever (DHF) patients, in samples collected up to five days after the onset of symptoms. Lines denote median values. * = $p < 0.05$, in Bonferroni's post hoc test.

secondary DHF (8.68 log₁₀ RNA copies/mL) infections was statistically significant ($p < 0.05$).

DISCUSSION

In this study we compared clinical and immunological parameters in patients from northeastern of Brazil with DF and DHF. Our results show a higher incidence of DHF in adult patients (≥ 15 years old), and the development of DHF has no correlation with secondary infection, which differs to the dogma of the epidemiology for DENV infection, and this may be an evidence that the pathogenesis of dengue disease can also be influenced by virus and host factors (Pozo-Aguilar et al. 2014).

We also found that platelet counts are significantly lower in DHF than in DF patients, in agreement with the findings of Alonzo et al. (2012). Despite the efforts, the mechanism of thrombocytopenia development in DHF patients is still poorly understood. IFN- β , an important anti-virus cytokine, was described to affect platelet production in vitro (Pozner et al. 2010). Therefore, high levels of IFN- β in primary DENV infections could contribute to platelets dysfunction and coagulation disorders, highlighting a possible role for IFN- β in the immunopathogenesis of severe dengue.

Several studies highlighted the role of type I IFN in virus infection control (Diamond et al. 2000), other studies suggest that DENV infection would be associated with an increase in IFN-I expression, in vitro and in vivo (Reis et al. 2007, Becquart et al. 2010). Hernández et al. (2014) observed that, in the early stages of DENV infection, levels of IFN- α were higher in DF than in DHF patients and this early strong interferon response with better clinical outcome. Otherwise, we could not find differences in the levels of circulating IFN- α when comparing DF with DHF patients during the acute phase of infection, and this difference could be explained by the sensitivity of the methods employed to measure circulating IFN- α , or even by the virulence of the different DENV serotypes studied. Similar results were obtained by Libraty et al. (2002) regarding IFN- α in DF and DHF during secondary DENV-3 infection. We did not find a correlation between IFN- α subtype 1 (IFN- α 1) mRNA levels and the concentration of circulating IFN- α . Since the quantitative RT-PCR carried out in our study only measured IFN- α 1 mRNA subtype, the ELISA assay instead was able to quantify all 13 different subtypes of circulating IFN- α , and this could have led to this discrepancy. The importance of IFN- β for the control of dengue infection was also demonstrated. Chen et al. (2013), no-

ticed that interferon regulatory factors (IRF) 3 and 7 deficient mice were not able to control DENV replication, as consequence these animals presented a high DENV burden in different tissues. Our report presents, for the first time, the primary DHF patients presented much higher levels of circulating IFN- β than in secondary DHF or DF ones. However, other studies reported that DF and DHF patients expressed similar levels of IFN- β (Torres et al. 2015) in primary or secondary infections (Tang et al. 2010), and the reason for these differences could lie in the genetic background of different studied populations, or in the sensitivity of the tests, as well as in DENV serotypes differences.

Elevation in AST and ALT levels has been associated with dengue illness progression (Rathakrishnan et al. 2012, Ferreira et al. 2015), evidencing liver compromising during dengue infection. We also found both AST and ALT levels elevated, especially in DHF patients, however no association was found between type-I IFN circulating levels with liver transaminases.

DENV has developed different strategies to subvert the host immune response (Muñoz-Jordan et al. 2003, Aguirre et al. 2012), what would allow a more efficient virus replication evidenced by higher viraemia, what could be associated with severe dengue forms. In fact, independent studies demonstrated that DHF patients presented higher circulating levels of DENV RNA than DF patients (Vaughn et al. 2000, Libraty et al. 2002, Guilarde et al. 2008). Our results showed that secondary infected DENV patients displayed higher levels of median virus load than primary ones. However, the small number of secondary DHF patients examined, makes the strength of this observation limited.

DF patients seems to secrete suitable levels of type I IFN in primary or secondary infections, which would ensure a proper anti-viral immune response, evidenced by lower levels of circulating virus. In the other hand, DHF seems to be a condition generated by an unbalanced immune response, which compromise the virus replication control. The results of this study suggest that DENV-3 infection might elicit a strong innate immune response characterised by higher levels of IFN- β in primary DHF patients from northeast of Brazil, which could be associated with development of severe dengue on this population. The severity of infection observed in our cohort could be a consequence of the association of clinical features (low platelet counts, high viraemia, bleeding manifestation) and immunity-related factors such as high levels of IFN- β . These findings may contribute to a better understanding of dengue pathogenesis.

ACKNOWLEDGEMENTS

To Dr Hen Phon-Too, from National University of Singapore, for kindly provide the Real Time PCR kit for dengue.

REFERENCES

- Aguirre S, Maestre AM, Pagni S, Patel JR, Savage T, Gutman D, et al. DENV inhibits type I IFN production in infected cells by cleaving human STING. *PLoS Pathog.* 2012; 8(10): e1002934.
- Alonzo MT, Lacuesta TL, Dimaano EM, Kurosu T, Suarez LA, Mapua CA, et al. Platelet apoptosis and apoptotic platelet clearance by macrophages in secondary dengue virus infections. *J Infect Dis.* 2012; 205(8): 1321-9.
- Applied Biosystems [Internet]. Amplification efficiency of TaqManR Gene Expression Assays. Assays tested extensively for qPCR efficiency. 2012. Available from: http://www3.appliedbiosystems.com/cms/groups/mcb_marketing/documents/generaldocuments/cms_040377.pdf.
- Applied Biosystems [Internet]. User Bulletin. Applied Biosystems 7500/7500 Fast Real-Time PCR Systems. 2010. Available from: http://www3.appliedbiosystems.com/cms/groups/mcb_support/documents/generaldocuments/cms_050637.pdf.
- Ashour J, Laurent-Rolle M, Shi PY, Garcia-Sastre A. NS5 of dengue virus mediates STAT2 binding and degradation. *J Virol.* 2009; 83(11): 5408-18.
- Assunção-Miranda I, Amaral FA, Bozza FA, Fagundes CT, Sousa LP, Souza DG, et al. Contribution of macrophage migration inhibitory factor to the pathogenesis of dengue virus infection. *Faseb J.* 2010; 24(1): 218-28.
- Back AT, Lundkvist A. Dengue viruses - an overview. *Infect Ecol Epidemiol.* 2013; 3: 19839.
- Becquart P, Wauquier N, Nkoghe D, Ndjoiy-Mbiguino A, Padilla C, Souris M, et al. Acute dengue virus 2 infection in Gabonese patients is associated with an early innate immune response, including strong interferon alpha production. *BMC Infect Dis.* 2010; 10: 356.
- Bozza FA, Cruz OG, Zagne SM, Azeredo EL, Nogueira RM, Assis EF, et al. Multiplex cytokine profile from dengue patients: MIP-1beta and IFN-gamma as predictive factors for severity. *BMC Infect Dis.* 2008; 8: 86.
- Chen HW, King K, Tu J, Sanchez M, Luster AD, Shresta S. The roles of IRF-3 and IRF-7 in innate antiviral immunity against dengue virus. *J Immunol.* 2013; 191(8): 4194-201.
- Chen LC, Lei HY, Liu CC, Shiesh SC, Chen SH, Liu HS, et al. Correlation of serum levels of macrophage migration inhibitory factor with disease severity and clinical outcome in dengue patients. *Am J Trop Med Hyg.* 2006; 74(1): 142-7.
- Cordeiro MT, Schatzmayr HG, Nogueira RM, Oliveira VF, Melo WT, Carvalho EF. Dengue and dengue hemorrhagic fever in the state of Pernambuco, 1995-2006. *Rev Soc Bras Med Trop.* 2007a; 40(6): 605-11.
- Cordeiro MT, Silva AM, Brito CA, Nascimento EJ, Magalhães MC, Guimarães GF, et al. Characterization of a dengue patient cohort in Recife, Brazil. *Am J Trop Med Hyg.* 2007b; 77(6): 1128-34.
- Diamond MS, Roberts TG, Edgil D, Lu B, Ernst J, Harris E. Modulation of dengue virus infection in human cells by alpha, beta, and gamma interferons. *J Virol.* 2000; 74(11): 4957-66.
- Dong T, Moran E, Chau NV, Simmons C, Luhn K, Peng Y, et al. High pro-inflammatory cytokine secretion and loss of high avidity cross-reactive cytotoxic T-cells during the course of secondary dengue virus infection. *PLoS ONE.* 2007; 2(12): e1192.
- Ferreira RA, de Oliveira SA, Gandini M, Ferreira LC, Correa G, Abiraude FM, et al. Circulating cytokines and chemokines associated with plasma leakage and hepatic dysfunction in Brazilian children with dengue fever. *Acta Trop.* 2015; 149: 138-47.
- Fink J, Gu F, Ling L, Tolfvenstam T, Olfat F, Chin KC, et al. Host gene expression profiling of dengue virus infection in cell lines and patients. *PLoS Negl Trop Dis.* 2007; 1(2): e86.
- Gandini M, Gras C, Azeredo EL, Pinto LM, Smith N, Despres P, et al. Dengue virus activates membrane TRAIL relocalization and IFN-alpha production by human plasmacytoid dendritic cells in vitro and in vivo. *PLoS Negl Trop Dis.* 2013; 7(6): e2257.

- Green S, Rothman A. Immunopathological mechanisms in dengue and dengue hemorrhagic fever. *Curr Opin Infect Dis*. 2006; 19(5): 429-36.
- Guilarte AO, Turchi MD, Siqueira Jr JB, Feres VC, Rocha B, Levi JE, et al. Dengue and dengue hemorrhagic fever among adults: clinical outcomes related to viremia, serotypes, and antibody response. *J Infect Dis*. 2008; 197(6): 817-24.
- Halstead SB, O'Rourke EJ. Dengue viruses and mononuclear phagocytes. I. Infection enhancement by non-neutralizing antibody. *J Exp Med*. 1977; 146(1): 201-17.
- Halstead SB. Pathogenesis of dengue: challenges to molecular biology. *Science*. 1988; 239(4839): 476-81.
- Hernández SIC, Puerta-Guardo H, Flores-Aguilar H, González-Mateos S, López-Martínez I, Ortiz-Navarrete V, et al. A strong interferon response correlates with a milder dengue clinical condition. *J Clin Virol*. 2014; 60(3): 196-9.
- Jones M, Davidson A, Hibbert L, Gruenwald P, Schlaak J, Ball S, et al. Dengue virus inhibits alpha interferon signaling by reducing STAT2 expression. *J Virol*. 2005; 79(9): 5414-20.
- Kurane I. Dengue hemorrhagic fever with special emphasis on immunopathogenesis. *Comp Immunol Microbiol Infect Dis*. 2007; 30(5-6): 329-40.
- Lee YR, Liu MT, Lei HY, Liu CC, Wu JM, Tung YC, et al. MCP-1, a highly expressed chemokine in dengue haemorrhagic fever/dengue shock syndrome patients, may cause permeability change, possibly through reduced tight junctions of vascular endothelium cells. *J Gen Virol*. 2006; 87(Pt 12): 3623-30.
- Li Y, Xie J, Wu S, Xia J, Zhang P, Liu C, et al. Protein kinase regulated by dsRNA downregulates the interferon production in dengue virus- and dsRNA-stimulated human lung epithelial cells. *PLoS ONE*. 2013; 8(1): e55108.
- Liang Z, Wu S, Li Y, He L, Wu M, Jiang L, et al. Activation of Toll-like receptor 3 impairs the dengue virus serotype 2 replication through induction of IFN-beta in cultured hepatoma cells. *PLoS ONE*. 2011; 6(8): e23346.
- Libraty DH, Endy TP, Hough HS, Green S, Kalayanarooj S, Suntayakorn S, et al. Differing influences of virus burden and immune activation on disease severity in secondary dengue-3 virus infections. *J Infect Dis*. 2002; 185(9): 1213-21.
- Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods*. 2001; 25(4): 402-8.
- Martina BE, Koraka P, Osterhaus AD. Dengue virus pathogenesis: an integrated view. *Clin Microbiol Rev*. 2009; 22(4): 564-81.
- Messina JP, Brady OJ, Pigott DM, Golding N, Kraemer MU, Scott TW, et al. The many projected futures of dengue. *Nat Rev Microbiol*. 2015; 13(4): 230-9.
- Muñoz-Jordan JL, Sánchez-Burgos GG, Laurent-Rolle M, García-Sastre A. Inhibition of interferon signaling by dengue virus. *Proc Natl Acad Sci USA*. 2003; 100(24): 14333-8.
- Nguyen TH, Lei HY, Nguyen TL, Lin YS, Huang KJ, Le BL, et al. Dengue hemorrhagic fever in infants: a study of clinical and cytokine profiles. *J Infect Dis*. 2004; 189(2): 221-32.
- Pozner FG, Ure AE, de Giusti CJ, D'Artri LP, Italiano JE, Torres O, et al. Junin Virus infection of human hematopoietic progenitors impairs in vitro proplatelet formation and platelet release via a bystander effect involving type I IFN signaling. *PLoS Pathog*. 2010; 6(4): 1-14.
- Pozo-Aguilar JO, Monroy-Martínez V, Díaz D, Barrios-Palacios J, Ramos C, Ulloa-García A, et al. Evaluation of host and viral factors associated with severe dengue based on the 2009 WHO classification. *Parasit Vectors*. 2014; 7: 590.
- Priyadarshini D, Gadia RR, Tripathy A, Gurukumar KR, Bhagat A, Patwardhan S, et al. Clinical findings and pro-inflammatory cytokines in dengue patients in Western India: a facility-based study. *PLoS ONE*. 2010; 5(1): e8709.
- Rathakrishnan A, Wang SM, Hu Y, Khan AM, Ponnampalavanar S, Lum LC, et al. Cytokine expression profile of dengue patients at different phases of illness. *PLoS ONE*. 2012; 7(12): e52215.
- Reis SRNI, Sampaio ALF, Henriques MGM, Gandini M, Azeredo EL, Kubelka CF. An in vitro model for dengue virus infection that exhibits human monocyte infection, multiple cytokine production and dexamethasone immunomodulation. *Mem Inst Oswaldo Cruz*. 2007; 102(8): 983-90.
- Ross TM. Dengue virus. *Clin Lab Med*. 2010; 30(1): 149-60.
- Sadler AJ, Williams BR. Interferon-inducible antiviral effectors. *Nat Rev Immunol*. 2008; 8(7): 559-68.
- Samuel CE. Antiviral actions of interferons. *Clin Microbiol Rev*. 2001; 14(4): 778-809.
- Sen GC. Viruses and interferons. *Annu Rev Microbiol*. 2001; 55: 255-81.
- Siegal FP, Kadowaki N, Shodell M, Fitzgerald-Bocarsly PA, Shah K, Ho S, et al. The nature of the principal type 1 interferon-producing cells in human blood. *Science*. 1999; 284(5421): 1835-7.
- Tang Y, Kou Z, Zhang F, Yao X, Liu S, Ma J, et al. Both viremia and cytokine levels associate with the lack of severe disease in secondary dengue 1 infection among adult Chinese patients. *PLoS ONE*. 2010; 5(12): e15631.
- Torres RE, Rivera RMC, Pino MAL, Burgos GGS. Serum levels of IFN-beta are associated with days of evolution but not with severity of dengue. *J Med Virol*. 2015; 88(3): 395-9.
- Umareddy I, Tang KF, Vasudevan SG, Devi S, Hibberd ML, Gu F. Dengue virus regulates type I interferon signalling in a strain-dependent manner in human cell lines. *J Gen Virol*. 2008; 89(Pt 12): 3052-62.
- Vaughn DW, Green S, Kalayanarooj S, Innis BL, Nimmannitya S, Suntayakorn S, et al. Dengue viremia titer, antibody response pattern, and virus serotype correlate with disease severity. *J Infect Dis*. 2000; 181(1): 2-9.
- WHO - World Health Organization [Internet]. Dengue: guidelines for diagnosis, treatment, prevention and control. 2009. Available from: <http://www.who.int/tdr/publications/documents/dengue-diagnosis.pdf>.
- WHO - World Health Organizations [Internet]. Dengue haemorrhagic fever: diagnosis, treatment, prevention, and control. 1997. Available from: <http://www.who.int/csr/resources/publications/dengue/Denguepublication/en/>.
- Wu S, He L, Li Y, Wang T, Feng L, Jiang L, et al. miR-146a facilitates replication of dengue virus by dampening interferon induction by targeting TRAF6. *J Infect*. 2013; 67(4): 329-41.