

Original Research Article

Identification of metabolites of kurarinone from *Sophora flavescens* Ait in rat urine by ultra-performance liquid chromatography with linear ion trap orbitrap mass spectrometry

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Abstract

Purpose: To study the *in vivo* metabolism of kurarinone, a flavandulyl flavanone which is a major constituent of Kushen and a marker compound with many biological activities, using ultra-performance liquid chromatography coupled with linear ion trap Orbitrap mass spectrometry (UPLC-LTQ-Orbitrap-MS).

Methods: Six male Sprague-Dawley rats were randomly divided into two groups. First, kurarinone was suspended in 0.5 % carboxymethylcellulose sodium (CMC-Na) aqueous solution, and was given to rats ($n = 3$, 2 mL for each rat) orally at 50 mg/kg. A 2 mL aliquot of 0.5 % CMC-Na aqueous solution was administered to the rats in the control group. Next, urine samples were collected over 0-24 h after the oral administrations and all urine samples were pretreated by a solid phase extraction (SPE) method. Finally, all samples were analyzed by a UPLC-LTQ-Orbitrap mass spectrometry coupled with an electrospray ionization source (ESI) that was operated in the negative ionization mode.

Results: A total of 11 metabolites, including the parent drug and 10 phase II metabolites in rat urine, were first detected and interpreted based on accurate mass measurement, fragment ions, and chromatographic retention times. The results were based on the assumption that kurarinone glucuronidation was the dominant metabolite that was excreted in rat urine.

Conclusion: The results from this work indicate that kurarinone *in vivo* is typically transformed to non-toxic glucuronidation metabolites, and these findings may help to characterize the metabolic profile of kurarinone.

Keywords: Kurarinone, Metabolites, *Sophora flavescens* Ait., Glucuronidation metabolites

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INTRODUCTION

Sophora flavescens Ait (Leguminosae) is a perennial shrub that is widely distributed in Asia, Oceania, and the Pacific Islands. For thousands of years, the dried roots of *Sophora flavescens*,

also named 'Kushen', have been traditionally used in East Asian countries as an herbal medicine for the treatment of hepatitis, gastrointestinal hemorrhage and skin diseases.

Phytochemical studies of *Sophora flavescens* described the isolation of quinolizidine alkaloids,

flavonoids and triterpenoids. The alkaloids in *Sophora flavescens* mainly consist of oxymatrine and matrine, which have been used clinically in China for the treatment of chronic hepatitis. Furthermore, several pharmacological products, such as injection oxymatrine and capsule matrine, are extensively used for the treatment of hepatitis and cancers in China. The flavonoids, which have been termed Kushen flavonoids, usually carry one or multiple prenyl units. Several studies have demonstrated that prenyl chains are crucial for the biological activity of flavonoids [1]. Therefore, the pharmacology of Kushen prenylated flavonoids has become an area of increased research because of their anti-diabetic, anti-arthritic, anti-inflammation, and anti-bacterial activities.

Kurarinone, a lavandulyl flavanone, is a major constituent of Kushen flavonoids, and it has a concentration range of 4.79-16.07 mg/g [2]. It is considered to be a marker compound with many biological activities, such as anti-immune [3], anti-bacterial [4], anti-atherosclerosis [5], anti-cancer [6], anti-carcinogenic [7] activities. Furthermore, some studies have suggested that kurarinone in combination with TRAIL has potential application for the treatment of tumors and gastric cancer [8], and it can also potentially increase the antitumor activity of Taxol [9].

Although kurarinone has various pharmacological effects, studies on the *in vivo* metabolism of kurarinone have rarely been reported. Yu [10] demonstrated that kurarinone induced remarkable cytotoxicity in primary rat hepatocytes and HL - 7702 cells, with IC_{50} values of 29.9 and 48.2 μ M, respectively. The metabolism of kurarinone by human liver microsomes *in vitro* indicated that kurarinone could be extensively glucuronidated to decrease its cytotoxicity [11].

Metabolism, which plays an important role in pharmacokinetics, can influence the biological activity and toxicity of drugs [12]. Further studies of kurarinone metabolism and the role of metabolism in biological activity or toxicity should be performed to provide a comprehensive overview of the clinical use of kurarinone.

EXPERIMENTAL

Isolation and identification of kurarinone

Kurarinone (Figure 1) was isolated in our laboratory from the ethanol extract of *Sophora flavescens* Ait.. A 5 kg quantity of dry kushen roots was extracted three times with ethanol. The ethanol extract was evaporated to dryness, and

the dry residue was suspended in water and further partitioned in succession with ethyl acetate and butanol. Part of the ethyl acetate extractives was chromatographed over 1000 g of a silica gel column (200 - 300 mesh) using a CH_2Cl_2 - MeOH step gradient elution, yielding ten fractions. The fractions (NO. 6) were chromatographed on MCI GEL CHP20P using a MeOH - H_2O step gradient elution, and they were purified further by preparative HPLC (C18) to yield kurarinone (3.01 g). Its structure was elucidated by the comprehensive interpretation of the ESI - MS and 1H , ^{13}C NMR data. The kurarinone purity was higher than 99 % according to HPLC - UV analysis.

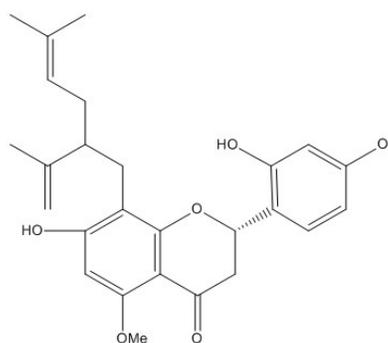


Figure 1: Chemical structures of kurarinone

Chemicals and materials

HPLC grade acetonitrile, methanol, and formic acid were purchased from Fisher (NJ, USA). Ultra-pure water was prepared using a Milli-Q water purification system (Millipore, Billerica, MA, USA). Waters Oasis HLB solid-phase extraction (SPE) cartridges (3 cc/60 mg, 30 μ m) were used. All other chemicals were of analytical grade and they were commercially available.

Animal experiments and drug administration

Six male Sprague-Dawley rats (200 \pm 10 g) were provided by Southern Medical University Experimental Animals Center (Guangzhou, China). Sprague-Dawley rats were housed in controlled environmental conditions (temperature: 24 \pm 2 $^{\circ}C$; relative humidity: 70 \pm 5 %). They were housed individually for a week for acclimatization, and they were fed ad libitum with standard laboratory chow (Southern Medical University Experimental Animals Center, Guangzhou China) and water. All the rats were fasted for more than 12 h with free access to water before the experiment. The rats were randomly divided into two groups: Group A (n = 3), drug group for urine; Group B (n = 3), control group for blank urine. All experimental protocols involving animals and their care were approved

(ref. no. SYXK (YUE) 2011 - 0015) by the institutional Animal Care and Use Committee at Southern Medical University (Guangzhou, China), and the experimental protocols followed the "Guide for the Care and Use of Laboratory Animals" [13].

Kurarinone (10 mg/mL) was suspended in 0.5 % carboxymethylcellulose sodium (CMC-Na) aqueous solution. The rats in the drug group were orally exposed to kurarinone at a dose of 50 mg/kg body weight. Next, a 2 mL aliquot of 0.5 % CMC-Na aqueous solution was administered to the rats in the control group. Urine samples were collected over a period of 0-24 h after the oral administrations, and all the biological samples from the same group were pooled into one sample.

Sample preparation

All the rat urine samples were pretreated by a solid-phase extraction (SPE) method. A pre-activated Oasis HLB solid phase extraction C18 column was loaded with a 1 mL urine sample, and then the cartridge was eluted by 5 mL water and 5 mL methanol successively. The methanol eluate was collected and evaporated to dryness by N_2 . The residues were reconstituted in 100 μ L methanol and then centrifuged at 14,000 rpm for 15 min at 4 °C. The final 2 μ L solution was injected into an UPLC-ESI-LTQ-Orbitrap MS for analysis.

Instruments and conditions

A Finnigan LTQ/Orbitrap MS (Thermo Electron, Bremen, Germany) coupled to an ESI source (Thermo Electron, Bremen, Germany) was used for all samples analysis. A Waters ACQUITY BEH C18 column (2.1 \times 100 mm i.d., 1.7 μ m) was used for separation of the metabolites at room temperature and a flow rate of 200 μ L/min. A linear gradient elution using 0.5 % formic acid aqueous solution (A) and acetonitrile (B) was applied with the following program: 0 - 3 min, 3 - 5 % B; 3 - 4 min, 5-30 % B; 4 - 20 min, 30 - 36 % B; 20 -21 min, 36 - 42 % B; 21 - 22 min, 42 - 60 % B; 22 - 25 min, 60 - 61 % B; 25 - 26 min, 61 - 80 % B; 26 - 30 min, 80 % B; 30 - 31 min, 80 - 3 % B, and 31 - 35 min, 3 % B.

An aliquot of 2 μ L was injected into the UPLC-LTQ Orbitrap MS. The optimized operating parameters in the negative ionization mode were as follows: capillary voltage of 35 V, electrospray voltage of 3.0 kV, capillary temperature of 350 °C, sheath gas flow rate of 30 (arbitrary units), auxiliary gas flow rate of 10 (arbitrary units), and tube lens voltage of 110 V.

Metabolites were detected by full scan mass-analysis from m/z 100 to 1000 at a resolving power of 30,000 with data-dependent MS² analysis triggered by the three most-abundant ions from the precursor list of predicted metabolites followed by MS³ analysis of the most abundant product ions. Collision-induced dissociation (CID) was performed with an isolation width of 2 Da. The collision energy was set to 35 %.

Peak selection and data processing

Thermo Xcalibur 2.1 workstation (Thermo Scientific, Bremen, Germany) was used for data acquisition and processing. To obtain the maximum number of fragments, the peaks detected with intensity > 10000 were selected for identification. The chemical formulas for all parent ions of the selected peaks were calculated from the accurate mass using a formula predictor by setting the parameters as follows: C [0 - 40], H [0 - 50], O [0 - 20], S [0 - 4], N [0 - 4], Cl [0 - 4], and ring double bond (RDB) equivalent value [0 - 15]. Other elements such as P and Br were not considered, as they were rarely present in the complex matrix. Blank biological samples were used as controls to compare with the analyte samples, and they were processed under the same conditions.

RESULTS

Fragmentation pathway of kurarinone

First, the MSⁿ fragmentation pattern of kurarinone was investigated to facilitate the structural identification of the metabolites. The parent ion showed a deprotonated ion $[M - H]^-$ at m/z 437.1950 (2.0 ppm, C₂₆H₂₈O₆) in negative ionization mode. The fragmentation of this parent ion provided a characteristic fragment ion at m/z 275 (C₁₇H₂₃O₃) by the loss of the C₉H₆O₃ moiety. The proposed fragmentation pattern of kurarinone is illustrated in Figure 2.

Metabolites of kurarinone in rats

After comparing the total ion chromatogram of the drug samples with the corresponding control samples, a total of 10 metabolites and the parent drug were detected and identified. The total ion chromatogram of the drug samples is shown in Figure 3. The chromatographic and mass spectrometric data of the parent drug and its metabolites are shown in Table 1.

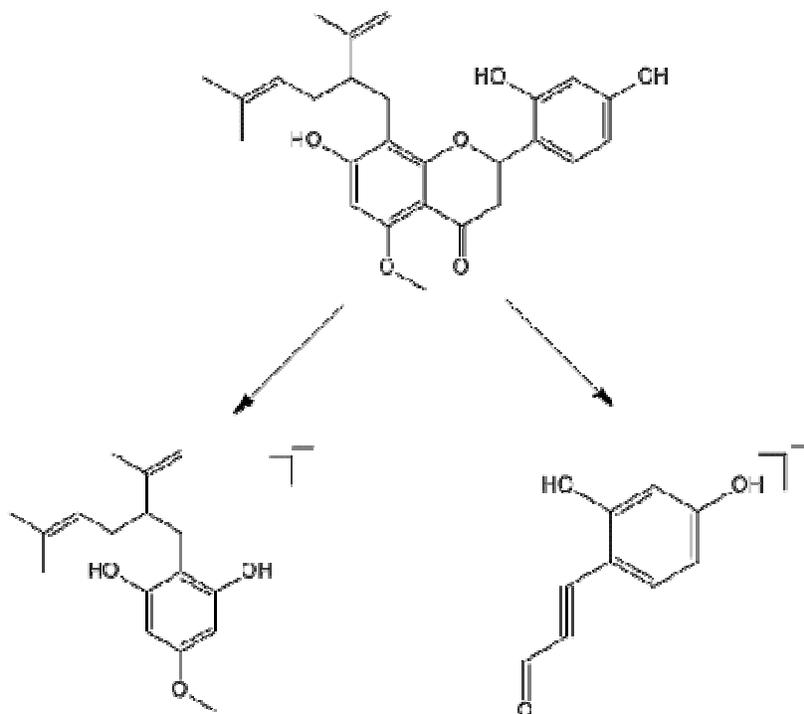


Figure 2: The proposed fragmentation pattern of kurarinone

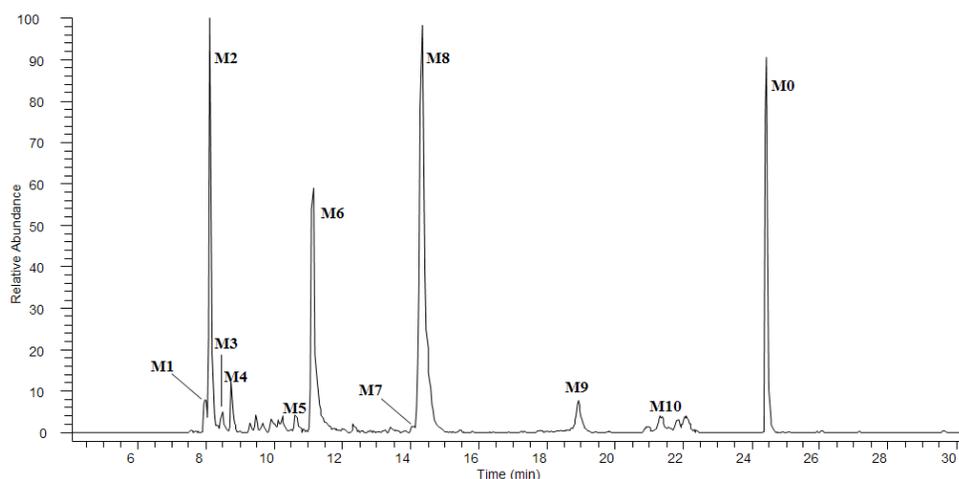


Figure 3: The total ion chromatograms of multiple metabolites in rats' urine

Metabolite M0

Metabolite M0 m/z 437.1958 (2.0 ppm, $C_{26}H_{29}O_6$) eluted at 24.42 min, and it was unambiguously identified as kurarinone by comparing the retention time, accurate mass measurement and fragment ions with the authentic reference material.

Metabolites M7 - 10

Metabolites M7, M8, M9, and M10 were eluted at 14.07, 14.33, 18.91 and 21.32 min, respectively. All of them yielded quasi-molecular ions of m/z

613.2269 (1.7 ppm, $C_{32}H_{37}O_{12}$), 613.2265 (2.4 ppm, $C_{32}H_{37}O_{12}$), 613.2267 (2.0 ppm, $C_{32}H_{37}O_{12}$), and 613.2268 (1.9 ppm, $C_{32}H_{37}O_{12}$), respectively. In the MS^2 spectra, there was an abundant product ion at m/z 437 that was produced by the neutral loss of 176 Da, indicating that they were glucuronide conjugates. Therefore, M7 - M10 were tentatively identified as glucuronidation of kurarinone.

Metabolites M5 - 6

Metabolites M5 and M6, with $[M - H]^-$ ions at m/z 789.2589 (1.4 ppm, $C_{38}H_{45}O_{18}$) and m/z

Table 1: Summary of kurarinone metabolites in rat urine

Peak	t _R	Theoretical mass <u>m/z</u>	Experimental mass <u>m/z</u>	Error (ppm)	Formula [M-H] ⁻	MS/MS fragment	Identification/reactions
1	7.97	629.2228	629.2218	-1.7	C ₃₂ H ₃₇ O ₁₃	MS ² [629]: 337 (100), 453(71), 291(38), 233(12)	Glucuronidation and Hydroxylation
2	8.11	629.2228	629.2219	-1.5	C ₃₂ H ₃₇ O ₁₃	MS ² [629]: 291 (100), 453(98), 337(92)	Glucuronidation and Hydroxylation
3	8.48	629.2228	629.2221	-1.2	C ₃₂ H ₃₇ O ₁₃	MS ² [629]: 453(100), 291(2)	Glucuronidation and Hydroxylation
4	8.74	629.2228	629.2217	-1.9	C ₃₂ H ₃₇ O ₁₃	MS ² [629]: 337 (100), 453(64), 233(12), 291(5)	Glucuronidation and Hydroxylation
5	10.6 2	789.2600	789.2589	-1.4	C ₃₈ H ₄₅ O ₁₈	MS ² [789]: 613 (100), 293(68), 437(10)	Diglucuronidation
6	11.1 4	789.2600	789.2588	-1.6	C ₃₈ H ₄₅ O ₁₈	MS ² [789]: 613 (100), 437(8)	Diglucuronidation
7	14.0 7	613.2279	613.2269	-1.7	C ₃₂ H ₃₇ O ₁₂	MS ² [613]: 337 (100), 437(92), 275(43), 419(21)	Glucuronidation
8	14.3 3	613.2279	613.2265	-2.4	C ₃₂ H ₃₇ O ₁₂	MS ² [613]: 337 (100), 437(85), 275(36), 419(14)	Glucuronidation
9	18.9 1	613.2279	613.2267	-2.0	C ₃₂ H ₃₇ O ₁₂	MS ² [613]: 437 (100), 293(78), 275(22), 419(8)	Glucuronidation
10	21.3 2	613.2279	613.2268	-1.9	C ₃₂ H ₃₇ O ₁₂	MS ² [613]: 437 (100), 293(68), 275(35), 419(4)	Glucuronidation
0	24.4 2	437.1958	437.1950	-2.0	C ₂₆ H ₂₉ O ₆	MS ² [437]: 161 (100), 275(28)	Parent drug

789.2588 (1.6 ppm, C₃₈H₄₅O₁₈) eluted at 10.62 and 11.14 min, respectively.

In MS² spectra, there was an abundant product ion at *m/z* 613 that was generated by the neutral loss of 176 Da, indicating that they were glucuronide conjugates. The MS³ of M5 and M6 possessed the same MS² spectra as M7 - 10, suggesting that the core structure of M5 and M6 was the di - glucuronidation of kurarinone.

Metabolites M1-4

Metabolites M1, M2, M3, and M4 eluted at 7.97, 8.11, 8.48, and 8.74 min, respectively. All of them yielded [M-H]⁻ ions at *m/z* 629.2218 (1.7 ppm, C₃₂H₃₇O₁₃), 629.2219 (1.5 ppm, C₃₂H₃₇O₁₃), 629.2221 (1.2 ppm, C₃₂H₃₇O₁₃), and 629.2217 (1.9 ppm, C₃₂H₃₇O₁₃), respectively. In the MS² spectra, there was an abundant product ion at *m/z* 453 that was produced by the neutral loss of 176 Da, indicating that they were glucuronide conjugates. The fragment ion at *m/z* 453 was present in the MS spectra, which was contained more O atoms more than M0 suggesting that it was a hydroxylation kurarinone ion. Therefore, M1 - M4 were tentatively identified as hydroxylation and glucuronidation of kurarinone.

DISCUSSION

Our results indicated that *in vivo* the kurarinone is typically transformed to non - toxic glucuronidation metabolites by phase II drug-metabolizing enzymes. In general, xenobiotics or drug compounds that are introduced into the body typically undergo metabolism, elimination and/or detoxification, then they are transformed to non-toxic metabolites by phase I and phase II drug-metabolizing enzymes. In general, conjugation with phase II DMEs (including sulfotransferases, UDP - glucuronosyltransferases and epoxide hydrolases) increases hydrophilicity, thereby enhancing excretion in the bile and/or the urine which consequently has a detoxification effect.

Although our results indicated that the UDP - glucuronosyltransferases (UGT) play important roles in kurarinone metabolic pathways and a series of glucuronidation metabolic reactions after the oral administration kurarinone, metabolites M1 - 4 indicated that the hydroxylation of kurarinone also occurs, but it has a short half - life. The *in vitro* investigation of kurarinone metabolism in human liver microsomes also suggests that the hydroxylation of kurarinone was much slower than

glucuronidation [11]. Our results suggested that kurarinone glucuronidation is considered to be more important than other metabolic pathway of kurarinone, and it is the major metabolic pathway *in vivo*.

CONCLUSION

The UPLC-LTQ-Orbitrap technique has been successfully applied to analyze the metabolic profile of orally administered kurarinone in Sparague-Dawley rat urine. A total of 10 metabolites and the parent drug were detected and identified based on accurate mass measurements, fragmentation patterns and chromatographic retention times.

To the best of our knowledge, this report is the first to describe the metabolism of kurarinone *in vivo*. The results indicate that kurarinone *in vivo* is typically transformed to non-toxic glucuronidation metabolites, and these findings help to characterize the metabolic profile of kurarinone in human liver microsomes *in vitro* by phase II drug-metabolizing enzymes.

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DECLARATIONS

Conflict of Interest

No conflict of interest associated with this work.

Contribution of Authors

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

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