



Research Article

Investigation Trp64Arg polymorphism of the beta 3-adrenergic receptor gene in nonobese women with polycystic ovarian syndrome

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Abstract

Background: Polycystic ovary syndrome (PCOS) is a multifactorial and heterogeneous disease that has a potent inheritable component based on familial clustering. Despite many studies in the genetic field of PCOS, the genes that are involved in the causes of this syndrome have not been thoroughly investigated.

Objective: The purpose of this study was to establish the occurrence of the Trp64Arg polymorphism of beta3 adrenergic receptor in non-obese women with PCOS.

Materials and Methods: This cross-sectional study was performed on 100 women with PCOS and normal women as the control group in Imam Khomeini Hospital of Tehran in 2016-2017. Peripheral blood sample (2 cc) was obtained from two groups for genomic DNA based on the gene bank. Polymorphisms were genotyped by of using ADRB3 Trp64Arg. Then the DNA was extracted by genomic kiagen kit. The primer was analyzed for PCR based on gene bank by using Primer3 software and then confirmed by primer Blast tool at NCBI site to conformity to the beta-3 adrenergic receptor gene. The protein changes were assessment by the Clastal W software.

Results: The sequence analysis presented in NCBI, transcript variant 1, with the code NM_000025.2, shows changes in the amino acid sequence of exon 1 in women with PCOS. Polymorphism in the codon 64 encoding the amino acid tryptophan (W) occurred in the nucleotide c.T190C, which changed the nucleotide T to C and then the amino acid sequence of the tryptophan was altered to arginine pW64R.

Conclusion: T-C polymorphism is evident in the codon 64 of the adrenergic β 3 receptor in patients with PCOS. Therefore, Beta3 adrenergic receptor gene polymorphism (Thr164lle) associates with this syndrome in nonobese women.

Key words: Codon 64, Beta-3 adrenergic receptor, Polymorphism, Polycystic ovarian syndrome.

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1. Introduction

Polycystic ovary syndrome (PCOS) has heterogeneous clinical characteristics: polycystic (PCOM), ovarian morphology hormone imbalance, and metabolic disorders such as insulin resistance (IR), diabetes, and obesity. These conditions are the cause of infertility ovarian and follicular maturation through disorders (1, 2). Years of experimental and human studies show that the overactivity of the sympathetic nervous system (SNS) can be the main cause for signs and complications in PCOS. The adrenergic β_3 receptor (ADR-B3) contains α1A, α1B, α1D, α 2A, α 2B, α 2C, β 1, and β 2 subgroups. These adrenoceptors are located in the adipose tissue, vascular endothelium, small intestine. The sympathetic activation of B3 is involved in cardio-inhibition, glucose uptake, lipolysis and relaxation of esophagus, bladder, and colon. The human β_3 receptor gene has been localized to chromosome 8 (8p12-8p11.1). B3ARs are also found in the brain, in areas such as the hypothalamus and the brain stem which can deliver multisynaptic innervation to white and brain adipose depots (3). SNS-mediated brown adipose tissue (BAT) thermogenesis activity diverse neurons found by these areas structures. SNS afferent neurons thermoregulatory pathways control as thermogenesis in BAT by thermoregulatory pathways with the interactions on the energy balance systems. The selective agonists of ADR-B3 can potently stimulate thermogenesis and lipolysis (4). Hadri and coworkers in 1997

showed that the transition from a fasted to a fed state in white and BAT in rats is associated with a decrease in \(\beta \)-adrenergic receptor mRNA levels and β3-adrenergic receptor responsiveness. They suggested that there is a close relationship between β3-adrenoceptor expression, plasma insulin, and food intake (5). There are many reports of the association of the ADR-B3 Trp64Arg gene polymorphism with obesity and metabolic syndrome (6-8). The gene encoding ADR-B2 polymorphism has been reported in hypertension (9), asthma (10), and autoimmune disease (11).

In this study, we investigated the association of Trp64Arg (rs 4994) in the first cytoplasmic region (Uniprot accession p13945) as one variant in the ADRB3 gene in PCOS.

2. Materials and Methods

2.1. Participants

This cross-sectional study was performed in November 2015-2016 on women with PCOs. In tota, 100 women with PCO participated from the Reproductive Health Research Center of Imam Hospital. All women were aged 20-40 years and the body mass index (BMI) <28 kg/m². They had no disease and no medication.PCOS diagnosis was according to the joint criteria of the European Society of Human Reproduction and Embryology and the American Society of Reproductive Medicine (ESHRE/ASRM) (12).

2.2. Sampling

In this study, 2cc of peripheral blood samples from the study (women with PCO) and control (normal) groups were collected in EDTA-treated tubes and froze at -20°C.

2.3. DNA extraction

The extraction of genomic DNA from all samples was done by using of the QIAamp DNA BIA Mini Kit Qiamp (Cat ID // 51106). The genomic DNA concentration and purity were assessed by using of UNICO-spectrophotometer (S2100SUV). Purity was determined using the standard A260/A280 and A260/A230 ratios

2.4. Primer design

2.4.1. Primers used to amplify exon 1 from the ADRB3 gene

Primer design was performed using Primer3 software for NG_011936.1 (Homo sapiens adrenoceptor beta 3 (ADRB3), RefSeqGene on chromosome 8) considering exon 1 and 2 regions in CDS region and then by using primer blast tool at NCBI site, it was approved in accordance with the ADRB3 gene exon 1 and 2 in NM_000025.2 without any mismatch with another region of the human genome. The fragment of interest was amplified with the PCR primers listed in Table I.

2.4.2. Genotyping of adrenoceptor beta 3 gene: The amplification of sample for preparation of PCR

25 μ L PCR mixture was prepared including: 20 ng of DNA, 10 pmol of each primer, 0.2 mmol/L of dNTPs, 2 mmol/L of MgCl2, and 1 U of Tag DNA polymerase. The thermal cycling situation for this study involved the original denaturation stage at 95°C for 10 min followed by 35 series of 95°C for 45 seconds, 59°C (exon 1) and 60°C (exon 2) for 1 minutes, and 72°C for 45 sec, and a final stage at 72°C for 10 minutes. All PCR products were exposed to electrophoresis in buffer of 1X TAE. The gel was marked with ethidium bromide (10mg/ml) and visualized under UV. The Gel documentation system was Transilluminator and photographed (Bio-Rad Laboratories) (Figures 1, and 2) and analysis after stored at 4°C (table I). So, the fragments obtained were subjected to electrophoresis in 1% agarose gel, and the size of the fragments and quality of them were checked (Figures 1, and 2). All samples were subjected to sequencing for genotyping and patient sequence analysis. The results were reproducible without any discrepancies. The PCR results from four samples were shown on 1% agarose gel for exon 1 and 2, respectively, as shown below.

The sequence provided for ADR β 3 in NCBI, transcript variant 1, with the code NM_000025.2 was used for investigation of changes in amino acid.

Table I. Primer used in amplification of ADRB3 gene

Gene (NG_011936)	Primer sequence (F, R, 5' 3')	Product length (base pairs)
Exon 1	F:TAGAGAAGATGGCCCAGGCT R:CGAGCCGTTGGCAAAGC	1323
Exon 2	F:GCTGGGTTGGAGTAGGGATG T:AGAGGTTGTGGAAAGGCTGG	1340

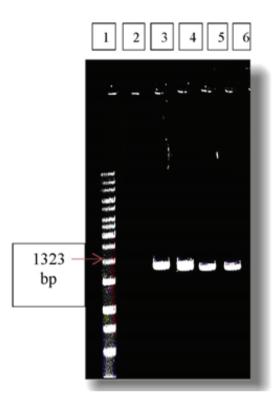


Figure 1. Representative agarose gel image showing amplification of exon 1 of ADRB3 gene by using its specific primers: (1) 100 bp DNA ladder; (2) Negative control of PCR; (3) Control sample 1; (4) Control sample 2; (5) Patient sample 3; and (6) Patient sample 4.

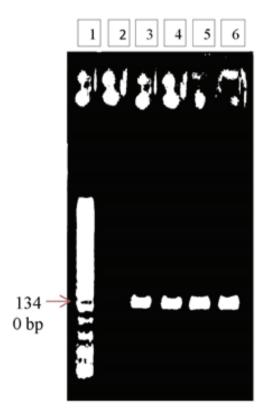


Figure 2. Representative agarose gel image showing amplification of exon 2 of ADRB3 gene by using its specific primers: (1) 100 bp DNA ladder; (2) Negative control of PCR; (3) Control sample 1; (4) Control sample 2; (5) Patient sample 3; (6) Patient sample 4.

2.5. Ethical consideration

This study was approved by the Ethical Committee of Tehran University of Medical Sciences (code: IR.TUMS.VCR.REC.2016.1329). The study points were explained to these women before they participated in the study, and informed consent was obtained from all.

2.6. Statistical analysis

IBM SPSS statistics (Statistical Package for the Social Sciences, version 24.0, SPSS Inc., and Chicago, IL) was used for analysis. To check the correlation between natural and mutated genotypes as well as homozygous and heterozygote genotypes Chi-square test (chi2) was used. P < 0.05 was considered as statistically significant level.

3. Results

3.1. PCR results

The quality and size of PCR products amplified for ADRB3 exon 1 and 2 sequences were analyzed using agarose gels. The comparison of PCR products with DNA size markers indicates that the PCR amplicons links to the expected PCR products. According to primer sequences (Table I), PCR product size

of 1323 and 1340 was confirmed (Figures 1, and 2) and their quality were good for sequencing.

3.2. Sequencing results

Our results show that codon 64 is associated with a polymorphism in women with PCOS. The point of mutation in PCR sequencing has been analyzed using Finch TV software in a woman with PCOS (Figure 3).

Our results show that there is a significant correlation between the presence of mutated genotype and PCOS. This result confirms that the mutagenic genotype can increases the chance of developing polycystic ovaries in women. OR: 2.546 (95% CI: 1.02-5.367) P: 0.012. The power of the study was also measured using the online GAS software (0.718), which was a significant measure that can indicate this study had high diagnostic power.

In this study, no significant correlation was found between homozygote and heterozygote genotype (p = 0.301), which is due to the high power of the study; it can be claimed that the sample size did not have an effect on statistical results. For complet, we used the Bootstrap test in which samples in each group were tested 1,000 times. Then, we used the Chi2 (it minimizes the sampling error) where these results had no difference (Table II).

Table II. Comparison of mutation and zygote in Beta3 adrenoceptor gene between women with PCOS and control group

Variables	Control	PCOS	Odd ratio (OR) (95%CI)	p-value	Statistical power
Wild mutant	68 (85%)	69 (69%)	2.546 (95%Cl:1.02-5.367)	0.012	
	12 (12%)	31 (31%)	,,		0.718
Homozygote heterozygote	68 (85%)	79 (21%)	1.506 (95%CI: 0.691-3.286)	0.301	5 // 10
	12 (15%)	21 (21%)			

Data presented as n (%); Chi-square



Figure 3. Sequencing of Codon64, c, mutant.

4. Discussion

In the present study, a significant association of Trp64Arg beta 3-adrenoceptor gene polymorphisms was observed in non-obese women with PCOS (BMI < 28 kg/m 2). Our result suggests that women with this mutation genotype have a significantly increased risk of PCOS (OR = 2.546, 95% CI: 1.02-5.367).

ADR- β agonists are potent activators of BAT in mice and humans (13). All sub-groups of Adrenoceptors β (β_{1-3}) have an active role in the metabolic processes like glycogenolysis, lipolysis, and insulin secretion. These adrenoceptors can cause metabolic syndrome (Mets) and PCOS. Familial and twin studies show that there is a genetic factor in women with PCOS with a polygenic pattern of heredity (14, 15). The studies on the PCOS exposure genes show that mostly these genes are associated with insulin sensitivity (16), sex hormone regulation (17), and metabolic disorders (18).

Energy metabolism can be a reliable predictor of weight gain. Overweight and obesity increase the incidence of PCOS and so the increasing of adiposity is considered a dominant characteristic in 40 to 60% of women with PCOS (19).

There are at least four metabolic parameters that can be predictive of weight gain: low metabolic rate, low fat oxidation rates, low spontaneous physical activity, and low activity of SNS.

The BAT, as a specific kind of adipose tissue with high activity, plays an important role in the metabolism of the body due to its high mitochondrial levels (20). Females with intact ovaries who are more insulin sensitive are protected from metabolic disorders because their BAT ratio is higher than the total fat (21-23). In an adult, Wang and coworkers showed that BAT is strongly activated by an increase in the circulating catecholamine concentration and there is a significant amount of BAT expressing β3-adrenergic receptor (23). Recent studies show that there is an important role of the ADR\$3 axis in human energy metabolism. This finding is confirmed by a common genetic codon of Trp64 \rightarrow Arg64 in ADR β 3, which can lead to metabolic abnormalities such as low levels of energy expenditure, high BMI (24), central obesity (25), hyperinsulinemia/IR (26), high blood pressure (BP) (27), type 2 diabetes (28), and even with gestational diabetes mellitus (29). These problems are also present in the symptoms and complications of women with PCO.

Studies of the last decade show that β 3-AR controls BAT thermogenesis in humans (30, 31). The SNS as the main regulator of BAT can be firing the SAS sub serving BAT for releasing of noradrenaline (NA) and activation of β 3 subtype (β 3-AR). This trigger activates the lipolysis process and mitochondrial uncoupling in BAT (31). A study on the BAT as a thermogenic machine, as well as the SNS-BAT axis, can help us to understand the complex etiopathology of Mets and PCOS.

ADR- B_3 (8p11.23) is essentially expressed in white/brown adipose tissue and is involved in thermogenesis thermogenesis and in the vascular endothelium for glucose uptake, lipolysis and cardio-inhibition/relaxation. The hypermethylation of this gene in visceral tissue leads to metabolic disorders (32).

In the present study, we investigated the Trp64Arg polymorphism of the beta 3-adrenergic receptor gene. For the selection of this codon, we must consider two important principles in women with PCOS as a dynamic study of two interwoven aspects: (1) epigenetic mechanisms and (2) neuroendocrine (autonomic/HPA/CRH imbalance) disorder.

Novel aetiopathological and treatment concepts can be raised from the fact that hypothalamic-pituitary-adrenal (HPA) axis dysfunction in PCOS like Mets could be initiated from sympathetic hyper activity in two syndromes. The homeostatic regulation of food intake is controlled by the HPA axis through many cross-links in the neural pathways of the neuroendocrine system (33). Firstly, it is the neurons of corticotrophin-releasing hormone (CRH) that contains the initial component of the HPA axis and are located in the paraventricular nucleus (PVN) of the hypothalamus, which is a major center for

controlling the feeding behavior (34). One of the other affective factors in BAT thermogenesis is the CRH, which is another important regulator of energy storage and adaptive thermogenesis. CRH acts via specific G-protein-coupled receptors. Real time RT-PCR and immunofluorescence confirmed that CRH (CRH-R1/ R2) can target in skeletal muscle and two types of adipose tissue: white adipose tissue (WAT) and BAT (35). CRH-R1/R2 receptors have been found in the stromal cells, thecal and follicular fluid in human ovaries. Ovarian CRH-like NA regulates steroidogenesis, follicular maturation, ovulation, and luteolysis (the so-called "aseptic" inflammatory) processes. In human, peripheral infusion of CRH stimulates fat oxidation and thermogenesis without changing lipolysis in adipose tissue or sympatho-adrenal activation. In normal female ovaries, there is no CRH in the oocyte of primordial follicle (36). But in the theca cells of womwn with PCO, CRH is associated with the rich of sympathetic nerves (37). In women with PCOS, the presence of CRH with its inhibitory role in oocyte maturation might be accompanied with the occurrence of follicle atresia that is observed in this syndrome (38). Zangeneh and coworkers in 2017 showed that hyponeurotrophinemia and decreasing CRH level in the serum of nonobese women with PCOS could reflect a deficiency of neuronal stress adaptation (39). This finding shows that central CRH and ovary CRH in women with PCOS is different from normal. Solinas and coworkers in 2006 for the first time reported the skeletal muscle's thermogenesis directly stimulates by CRH. They demonstrated that this thermogenic effect requires both phosphatidylinositol 3-kinase and AMP-activated protein kinase signaling, which can control the thermogenesis in this tissue by its protection against skeletal muscle lipotoxicity and IR (40). IR in women with PCO is not generalized in all tissues. In rat modeling of the knock-out of insulin receptors, hyperandrogenism and anovulation enhanced (41). Defects in metabolic function by insulin can occur in adipose and muscle tissues, but in the ovaries, mitogen and steroidogenic actions are maintained (42). Most women with PCOS have metabolic IR, partly due to obesity or due to the genetic predisposition. Ovarian insulin receptor can increases the response of theca cells to LH, and by increasing the expression of P450c17 and 3β-HSD2 leads to elevate of androgen production (43). In women with PCO, IR as a metabolic abnormality has a significant link with chronic inflammation, and polycystic ovarian syndrome is a low-level chronic inflammation disease. Many studies show that this inflammation has an essential role in the IR and metabolic consequences in women with PCOS (44, 45). The set point of body weight is the balance between energy intake and energy expenditure, and the central role of insulin axis in energy imbalance can contribute to the pathogenesis of obesity. Insulin as the mediator of feeding-related can increase thermogenesis. Metabolic activity of BAT can be activated by agonists of β 3-adrenoceptor, and NA transporter (NAT) can block it which is measurable using [18F] fluorodeoxyglucose positron emission tomography/computed tomography (PET/CT) in rat. By this method, Baranwal and coworkers showed that the beta-3-adrenergic agonist CL316, 243 has a potential role on BAT for modulating blood glucose levels (46).

Studies show that pharmacogenetics of β 3AD agonists and antagonists of NAT can help us to treat obesity and diabetes (47, 48).

All of these bodies of evidences more confirm that PCOS is a heterogeneous disease with metabolic disorders and on the base of our hypothesis; Trp64Arg polymorphism of beta3 adrenoceptor can increase the chance of PCOS.

5. Conclusion

The genomic studies help us for novel aetiopathological and treatment concepts of PCOS. T-C polymorphism is evident in the codon 64 of the ADR-B3 in patients with PCOS and Beta3 gene polymorphism (Thr164lle) associated with this syndrome. We need to study the future research that investigated the interactions of risk genotypes, environmental factors, and epigenetic encoding in the pathophysiology of PCOS.

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Conflict of Interest

The authors declare that there is no conflict of interest.

References

- [1] Franks S. Polycystic ovary syndrome. *N Engl J Med* 1995; 333: 853–861.
- [2] Dumesic DA, Padmanabhan V, Abbott DH. Polycystic ovary syndrome and oocyte developmental competence. *Obstet Gynecol Surv* 2008; 63: 39–48.
- [3] Richard JE, López-Ferreras L, Chanclón B, Eerola K, Micalle P, Skibicka KP, et al. CNS β_3 -adrenergic receptor activation regulates feeding behavior, white fat browning, and body weight. *Am J Physiol Endocrinol Metab* 2017; 313: E344–E358
- [4] Labbé SM, Caron A, Lanfray D, Monge-Rofarello B, Bartness TJ, Richard D. Hypothalamic control of brown adipose tissue thermogenesis. Front Syst Neurosci 2015; 9:150.
- [5] Hadri KE, Charon C, Pairault J, Hauguel-De Mouzon S, Quignard-Boulangé A, Fève B. Down-regulation of beta3adrenergic receptor expression in rat adipose tissue during the fasted/fed transition: evidence for a role of insulin. *Biochem J* 1997; 323: 359–364.
- [6] Zhang H, Wu J, Yu L. Association of Gln27Glu and Arg16Gly polymorphisms in Beta2-adrenergic receptor gene with obesity susceptibility: A meta-analysis. *PLoS One* 2014; 9: e100489.

- [7] Malik SG, Saraswati MR, Suastika K, Trimarsanto H, Oktavianthi S, Sudoyo H. Association of beta3-adrenergic receptor (ADRB3) Trp64Arg gene polymorphism with obesity and metabolic syndrome in the Balinese: a pilot study. BMC Res Notes 2011; 4: 167.
- [8] Weyer C, Gautier JF, Danforth E Jr. Development of beta 3-adrenoceptor agonists for the treatment of obesity and diabetes—an update. *Diabetes Metab* 1999; 25: 11–21.
- [9] Atia A, Abdullah A, Alrawaiq N. Overview of the role of B2-adrenergic receptor variants in human hypertension. *International Journal of PharmTech Research* 2014; 6: 1611–1615.
- [10] Reihsaus E, Innis M, MacIntyre N, Liggett SB. Mutations in the gene encoding for the beta 2-adrenergic receptor in normal and asthmatic subjects. Am J Respir Cell Mol Biol 1993; 8: 334–339.
- [11] Xu W, Liu Y, Ye D. Association between IL-33 gene polymorphisms (rs1929992, rs7044343) and systemic lupus erythematosus in a chinese han population. *Immunol Invest* 2016; 45: 575–583.
- [12] Rotterdam ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. Fertil Steril 2004; 81: 19–25.
- [13] Cypess AM, Weiner LS, Roberts-Toler C, Franquet Elía E, Kessler SH, Kahn PA, et al. Actvation of human brown adipose tissue by a β3-adrenergic receptor agonist. Cell Metab 2015; 21: 33–38.
- [14] Fratantonio E, Vicari E, Pafumi C, Calogero AE. Genetics of polycystic ovarian syndrome. Reprod Biomed Online 2005: 10: 713–720.
- [15] Jahanfar S, Eden JA. Genetic and non-genetic theories on the etiology of polycystic ovary Syndrome. Gynecol Endocronol 1996; 10: 357–364.
- [16] Obermayer-Pietsch B, Trummer C, Schwetz V, Schweighofer N, Pieber T. Genetics of insulin resistance in polycystic ovary syndrome. *Curr Opin Clin Nutr Metab Care* 2015; 18: 401–406.
- [17] Dadachanji R, Shaikh N, Mukherjee S. Genetic variants associated with hyperandrogenemia in pcos pathophysiology. *Genet Res Int* 2018; 2018: 7624932.
- [18] De Leo V, Musacchio MC, Cappelli V, Massaro MG, Morgante G, Petraglia F. Genetic, hormonal and metabolic aspects of PCOS: an update. *Reprod Biol Endocrinol* 2016; 14: 38
- [19] Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, et al. The androgen excess and PCOS society criteria for the polycystic ovary syndrome: the complete task force report. Fertil Steril 2009; 91: 456–488.
- [20] Bartelt A, Heeren J. Adipose tissue browning and metabolic health. Nat Rev Endocrinol 2014; 10: 24–36.
- [21] Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, et al. Identification and importance of brown adipose tissue in adult humans. N Engl J Med 2009; 360: 1509–1517.
- [22] Pfannenberg C, Werner MK, Ripkens S, Stef I, Deckert A, Schmadl M, et al. Impact of age on the relationships of brown adipose tissue with sex and adiposity in humans. *Diabetes* 2010; 59: 1789–1793.

- [23] Wang Q, Zhang M, Ning G, Gu W, Su T, Xu M, et al. Brown adipose tissue in humans is activated by elevated plasma catecholamines levels and is inversely related to central obesity. PLoS One 2011; 6: e21006.
- [24] Kurokawa N, Young EH, Oka Y, Satoh H, Wareham NJ, Sandhu MS, et al. The ADRB3 Trp64Arg variant and BMI: a meta-analysis of 44 833 individuals. *Int J Obes (Lond)* 2008; 32: 1240–1249.
- [25] Baturin AK, Pogozheva AV, Sorokina Elu, Makurina ON, Tutel'ian VA. The Trp64Arg polymorphism of beta3adrenoreceptor gene study in persons with overweight and obesity. Vopr Pitan 2012; 81: 23–27.
- [26] Nolsøe RL, Hamid YH, Pociot F, Paulsen S, Andersen KM, Borch-Johnsen K, et al. Association of a microsatellite in FASL to type II diabetes and of the FAS-670G>A genotype to insulin resistance. *Genes Immun* 2006; 7: 316–321.
- [27] Li YY, Lu XZ, Wang H, Zhou YH, Yang XX, Geng HY, et al. ADRB3 Gene Trp64Arg polymorphism and essential hypertension: A meta-analysis including 9,555 subjects. Front Genet 2018; 9: 106–115.
- [28] Ryuk JA, Zhang X, Ko BS, Daily JW, Park S. Association of β3-adrenergic receptor rs4994 polymorphisms with the risk of type 2 diabetes: A systematic review and metaanalysis. *Diabetes Res Clin Pract* 2017; 129: 86–96.
- [29] Guan L, Cui X, Zhou H. Meta-analysis of the association between the Trp64Arg polymorphism of the beta-3 adrenergic receptor and susceptibility to gestational diabetes mellitus. J Obstet Gynaecol 2018; 38: 172–176.
- [30] Tseng YH, Kokkotou E, Schulz TJ, Huang TL, Winnay JN, Taniguchi CM, et al. New role of bone morphogenetic protein 7 in brown adipogenesis and energy expenditure. *Nature* 2008; 454: 1000–1004.
- [31] Mund RA, Frishman WH. Brown adipose tissue thermogenesis: β 3-adrenoreceptors as a potential target for the treatment of obesity in humans. *Cardiol Rev* 2013; 21: 265–269.
- [32] Fujisawa T, Ikegami H, Yamato E, Takeawa K, Nakagawa Y, Hamada Y, et al. Association of Trp64Arg mutation of the β3-adrenergic-receptor with NIDDM and body weight gain. *Diabetologia* 1996; 39: 349–352.
- [33] Berthoud HR. Mind versus metabolism in the control of food intake and energy balance. *Physiol Behav* 2004; 81: 781–793.
- [34] Richardson RD, Omachi K, Kermani R, Woods SC. Intraventricular insulin potentiates the anorexic effect of corticotropin releasing hormone in rats. Am J Physiol Regul Integr Comp Physiol 2002; 283: R1321–R1326.
- [35] Lu B, Diz-Chaves Y, Markovic D, Contarino A, Penicaud L, Fanelli F, et al. The corticotrophin-releasing factor/urocortin system regulates white fat browning in mice through paracrine mechanisms. *Int J Obes (Lond)*. 2015; 39: 408–17.
- [36] Smith SR, de Jonge L, Pelleymounter M, Nguyen T, Harris R, York D, et al. Peripheral administration of human corticotropin-releasing hormone: a novel method to increase energy expenditure and fat oxidation in man. *J Clin Endocrinol Metab* 2001; 86: 1991–1998.
- [37] Mastorakos G, Karoutsou El, Mizamtsidi M. Corticotropin releasing hormone and the immune/inflammatory

- response. Eur J Endocrinol 2006; 155: S77-S84.
- [38] Kiapekou E, Zapanti E, Mastorakos G, Loutradis D. Update on the role of ovarian corticotropin-releasing hormone. *Ann NY Acad Sci* 2010; 1205: 225–229.
- [39] Zangeneh FZ, Naghizadeh MM, Bagheri M, Jafarabadi M. Are CRH & NGF as psychoneuroimmune regulators in women with polycystic ovary syndrome? *Gynecol Endocrinol* 2017; 33: 227–233.
- [40] Solinas G, Summermatter S, Mainieri D, Gubler M, Montani JP, Seydoux J, et al. Corticotropin-releasing hormone directly stimulates thermogenesis in skeletal muscle possibly through substrate cycling between de novo lipogenesis and lipid oxidation. *Endocrinology* 2006; 147: 31–38
- [41] Wu S, Divall S, Nwaopara A, Radovick S, Wondisford F, Ko C, et al. Obesity-induced infertility and hyperandrogenism are corrected by deletion of the insulin receptor in the ovarian theca cell. *Diabetes* 2014; 63: 1270–1282.
- [42] Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev* 2012; 33: 981– 1030.

- [43] Baptiste CG, Battista MC, Trottier A, Baillargeon JP. Insulin and hyperandrogenism in women with polycystic ovary syndrome. *J Steroid Biochem Mol Biol* 2010; 122: 42–52.
- [44] Flaa A, Aksnes TA, Kjeldsen SE, Eide I, Rostrup M. Increased sympathetic reactivity may predict insulin resistance: an 18-year follow-up study. *Metabolism* 2008; 57: 1422–1427.
- [45] Zafari Zangeneh F, Naghizadeh MM, Masoumi M. Polycystic ovary syndrome and circulating inflammatory markers. *Int J Reprod Biomed* 2017; 15: 375–382.
- [46] Baranwal A, Mirbolooki MR, Mukherjee J. Initial assessment of β 3-adrenoceptor-activated brown adipose tissue in streptozotocin-induced type 1 diabetes rodent model using [18F] fluorodeoxyglucose positron emission tomography/computed tomography. *Molecular Imaging* 2015; 14: 561–566.
- [47] Arch JR, Wilson S. Prospects for beta 3-adrenoceptor agonists in the treatment of obesity and diabetes. Int J Obes Relat Metab Disord 1996; 20: 191–199.
- [48] Sawa M, Harada H. Recent developments in the design of orally bioavailable beta3-adrenergic receptor agonists. *Curr Med Chem* 2006; 13: 25–37.